

# Effect of Gloriosa Superba Root Extract on Development of Chick Embryo

Shahista A. Shaikh<sup>\*1</sup>, Laxman D. Ghaywat<sup>2</sup>, Rohini Kanawade<sup>3</sup>

<sup>1</sup>Department Chemistry, S.M.B.S.T.College, Sangamner, Ahmednagar, Maharashtra, India <sup>2</sup>Department Zoology, S.M.B.S.T.College, Sangamner, Ahmednagar, Maharashtra, India <sup>3</sup>Department Biology, B.G.P Sahyadri Junior College, Sangamner, Ahmednagar, Maharashtra, India

## ABSTRACT

Gloriosa superba has been widely used for several medicinal purposes in Indian traditional medicine system. It is traditionally used for the treatment of gout, chronic ulcers, haemorrhoids, cancer, and leprosy and also for inducing labour pains etc. The seed and tubers of this plant contain cholchine as major constituent. The purpose of study is to evaluate the effect of Gloriosa superba root extract on development of chick embryo. The Soxhelt aqueous extract of tuber was extracted using methanol. The effect of different concentration of tuber extract was carried out on eggs. The effect of extract was found that it stops development of chick embryo. Gloriosa superba showed abortifacient and oxytocic activity activity due to the presence of colchicines.

Keywords: Abortificient ; Gloriosa superba; Oxytocic activity

## I. INTRODUCTION

In traditional medicinal culture thousand of references are available for the use of plants for the problems related to reproduction. Traditionally used herbs have great importantance in modern world because of their efficacy, safety and minimal side effect on the human health as compared to chemically synthesized medicines. Gloriosa superba Linn. (Family-Liliaceae) is a flowering plant commonly known as flame lily, fire lily, climbing lily. It is semi-woody, climbing using tendrils on bushes<sup>[1]</sup>. The plant has great importance in Ayurveda is used in inflammations<sup>[12]</sup>, and anthelmintic<sup>[5]</sup>, gout, rheumatoid arthritis, gonorrhea and relieving fever<sup>[2]</sup>. Root extract are used to cure leprosy, ulcer, piles, skin diseases and show anti-dote property against snake bite. It is also used as abortifacient<sup>[4]</sup> and extract of tuber are applied topically during childbirth for reducing labor pains<sup>[6][10]</sup>. The leaf powder is extensively used to overcome jaundice and head lice<sup>[10[7][8]</sup> anti-microbial<sup>[9]</sup>, antibacterial<sup>[11]</sup> The properties of tuber of the plant are reported<sup>[8]</sup>.

Literature study show that a plant seed and root are great sorce of colchicine and colchicoside. Colchicine is a powerful antimitotic agent that blocks or suppresses cell division by inhibiting mitosis, the division of a cell's nucleus. These phytochemical constituents are responsible for the plant's abortifacient and oxytocic activities<sup>[16][17]</sup>. Considering traditional importance of

Gloriosa *superba*, the aim of study was to investigate the effect of root extract of Gloriosa *superba* on embryonic development of chick egg.

## **II. MATERIAL AND METHODS**

#### Collection and authentication of the plant material

The tuber of the healthy plant Gloriosa *superba* was collected from Mordara, Pemgiri, Sangamner (Maharashtra). The plant material was taxonomically identified with the help of available literature.

## **Preparation of extract**

Freshly collected tubers were washed with distilled water. The cleaned tubers were subsequently dried under sunshade to remove moisture completely and powdered by using mechanical grinder. The powdered plant material was extracted using methanol with Soxhlet apparatus for 18 h. The extract was concentrated by evaporating on water bath and dried to obtain a dark brown semi-solid mass.

## Phytochemical screening

Identification of the phytochemical was carried out on the plant extract to find out the presence of alkaloids, steroids, proteins and glycosides by using specific reagents<sup>[18]</sup>.

#### Experiment

Chick Embryos: The chick embryos are easily available in large numbers hence chick embryo has been used to observe the teratological studies, because the postblastula chick embryo and the mammalian embryo are similar, and thus the chick embryo is a good model for studying vertebrate embryonic development. All aspects of animal care compiled with the ethical guidelines and technical requirements were approved by the Institutional Animal Ethics Committee (IAEC) and Institutional Review Board (IRB).

Eighteen (18 nos.) fertile, pathogen free eggs incubated at 35°C for 48 hrs. and 75% relative humidity until the embryos reached stage ten of development according to Hamburger and Hamilton [12]. All the eggs were labelled and divided into three groups consisting of six eggs per group. The Group one (G1)- Normal (uninjected) eggs, Group two (G2)- Injected with physiological saline, Group three (G3)- injected with the 5 mm root extract of Gloriosa *superba* and 5 mm physiological saline

#### Dosage of root extract of Gloriosa superba

Dosage was prepared by diluting 0.5 gm root extract of Gloriosa *superba* in 1 ml physiological saline(50 % solution).

## **Method of injection**

Eggs were wiped with 70% alcohol and labelled on the outer shell. A hole was made on the blunt pole of the egg with a sharp and thick needle under aseptic condition. Using a sterile needle and a syringe, 0.5 ml dosage of 50% solution of root extract in saline was injected to the corresponding groups of eggs. The gap created in the eggs was sealed.

All eggs were kept in incubator for 48 hrs.

## Observations

In each group, to determine the development of the chick embryos, the eggs were removed from the incubator after 48 hrs. The egg shell was opened to see the embryo. All the chick embryos were transferred to a petri dish by the careful sterile dissection.

On observation it is revealed that embryos from G1 and G2 shows development and embryo from G3 showed no development. (Table 1) (figure 1)

**Table 1.** Experimental design of chick embryos

Sr. No.	Groups	Observation
G1	Normal group	Normal
		Development
G2	Group injected with	Normal
	physiological saline	Development
G3	Group injected with	No Development
	dilute root extract of	
	Gloriosa <i>superba</i> in	
	physiological saline	



Figure 1. (I) chick embryo of Normal group, (II) chick embryo injected with physiological saline, (III) injected with dilute root extract of Gloriosa *superba* in physiological saline

## **III.** Conclusion

The experiment indicates that due to the injection of root extract, there is no further growth seen in chick embryo. It gives scientific evidence for the traditional use of Gloriosa *superba* as a abortifacient.

## **IV. Acknowledgment**

Authors are very much thankful to all articles and journals taken as a references, reviewed and discussion for this article. The authors are also grateful to the department of zoology for their technical support to complete this study.

## **V. REFERENCES**

- S Padmapriya1, K Rajamani, V A SathiyamurthyGlory Lily (Gloriosa superba L.) -A Review International Journal of Current Pharmaceutical Review and Research; 7(1); 43-49
- [2]. Nadakani AKIndian Materia Medica, 4th ed.: Popular Prakashan LimitedMumbai.2002:234-236.

- [3]. Kirthikar KR and Basu BDIndian medicinal plants2nd edL.MBasu Allahabad, Vol III, 1988, 1932-33.
- [4]. Kala, C., Farooquee N., and Dhar UPrioritization of medicinal plants on the basis of available knowledge,
- [5]. existing practices and use value status in Uttaranchal, IndiaBiodiversity and Conservation, 2004, 13(2): 453-469.
- [6]. Pawar B.M., et al.(2010)Anthelmintic activity of Gloriosa superba Linn (Liliaceae)International Journal of Pharma Tech Research 2(2) 1483-87
- [7]. Kirthikar KR and Basu BDIndian medicinal plants2nd edL.MBasu Allahabad, Vol III, 1988, 1932-33.
- [8]. Kurian JCPlant that heal, 7th edn, Vol-IGlory Lily, Oriental Watchman Publishing House, Pune, 2004:75.
- [9]. Kuldeep Gan U, Brij Bhushan and TiwariHepatoprotective activity of tuberous roots
- [10]. of Gloriosa superbaIndian J Nat Prod2007;23(4):8-12.
- [11]. Alagesaboopathi, C, Antimicrobial screening of selected medicinal plants in Tamilnadu, IndiaJournal of Microbiology, 2011, 5(6), 617-621.
- [12]. Lal, H.S and P.K.Mishra.(2011)Gloriosa superba –an endangered plant spotted for the first time from forest of Tpchanchi, Hazaribag (Jharkhand) IndiaScience Research Reporter 1(2) 61-64
- [13]. Rehana banu, and Nagarajan NAntibacterial Potential Of Glory Lily, Gloriosa Superba Linn, International Research Journal Of Pharmacy, 2011, 2(3), 139-142.
- [14]. Abhishek Mathur, Satish K Verma, Santosh K Singh, Deepika Mathur, Prasad GBKS, and Dua VK, Investigation Of Anti-Inflammatory Properties of Swertia Chirayta And Gloriosa Superba, Recent Research in Science and Technology, 2011, 3(3), 40-
- [15]. Senthilkumar MPhytochemical Screening and Antibacterial Activity of Gloriosa superba LinnInternational Journal of Pharmacognosy and Phytochemical Research 2013; 5(1); 31-36
- [16]. Haroon, K., Murad, A.Kand Iqbal HEnzyme inhibition activities of the extracts from rhizomes of Gloriosa superba Linn (Colchicaceae)Journal of

enzyme inhibition and medicinal chemistry, 2008, 22 (6) 722-725.

- [17]. Kokate, CK., Purohit, AP., and Gokhale,S.BPharmacognosy, Nirali Prakashan, Pune, 2004, 506
- [18]. Arati A Malpani, Urmila M Aswar, Shiv K Kushwaha, GN Zambare and SL BodhankarEffect of the Aqueous Extract of Gloriosa superba Linn (Langli) Roots on Reproductive System and Cardiovascular Parameters in Female RatsTropical Journal of Pharmaceutical Research April 2011; 10 (2): 169-176
- [19]. KPLatha, HKirana and HNGirish.Anti-Implantation Activity of the droalcoholic Tuber Extract of Gloriosa superba Linn in Female Albino RatsIJAPBC – Vol2(3), Jul-Sep, 2013 ISSN: 2277 – 4688.
- [20]. Khandelwal KRPractical Pharmacognosy, 18th edPune: Nirali Prakashan; 2007, pp15-18.