

Bacterial Blight Resistance in Rice: A Review

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ABSTRACT

Bacterial blight disease, caused by *Xanthomonas oryzaepv.oryzae*(Xoo), is one of the most serious diseases in rice producing areas. For combating the disease, the most effective and economical measure is exploitation of host plant resistance. To date, more than 38 R genes for BB resistance have been reported. Since the chemical control is not effective, the utilization of resistant varieties carrying resistance genes have been considered to be the most effective way to control the disease. With the development of a wide range of molecular techniques, marker assisted breeding is now used to enhance traditional breeding programs to improve crops. Among various PCR based markers, SSR markers are more popular in rice because they are highly informative, mostly mono locus, co-dominant, easily analyzed and cost effective. Besides disease management, bioagents also stimulate plant growth, even if there is no disease, which results in better yield. Antagonistic potential of different bioagents against bacterial leaf blight of rice has been reported by several workers.

Keywords: : Rice, Bacterial Blight Disease, Resistance Genes, SSR Markers

INTRODUCTION

Rice (*Oryza sativa* L.) (2n = 24) belonging to the family Poaceae is the staple food for one third of the world's population that occupies almost one-fifth of the total land area covered under cereals (Chakravarthi and Naravaneni, 2006). Bacterial blight (BB), caused by *Xanthomonas oryzaepv.oryzae* (Xoo), is a widely distributed and devastating diseases of both conventional and hybrid rice in south-eastern Asia (Mew, 1987; Nino-Liu et al., 2006). Bacterial blight disease is a systemic disease and can cause severe yield loss up to 50 % depending on growth stage, geographic localization and season (Gnanamanickam et al., 1999; Nino-Liu et al., 2006). In Taiwan, bacterial blight disease often occurs in the second crop season, and its annual incidence area is usually more than 20,000 hectares, accounting for approximately 4 % of the Taiwanese rice production area. Recently, this disease has become more and more serious because of climate change (Hsieh, 2003; Wang et al., 2013). At present, the prevention of bacterial blight includes field management, fertilizer control and resistance breeding. In practice, the cultivation of

resistant rice varieties has been proposed to be the most effective strategy to prevent bacterial blight disease (Khush et al., 1989; Shen and Ronald, 2002; Yang et al., 2003). The durable and broad resistance of plants was found to be usually governed by multiple genes or quantitative trait loci (QTLs) (Johnson, 1984). Therefore, the discovery of a resistance gene against Xoo is an important area of research leading to breeding programs. With the development of a wide range of molecular techniques, marker assisted breeding is now used to enhance traditional breeding programs to improve crops (Frey et al., 2004). Among various PCR based markers, SSR markers are more popular in rice because they are highly informative, mostly mono locus, co-dominant, easily analyzed and cost effective (Gracia et al., 2004). Simple Sequence Repeats (SSRs) or microsatellites are most suited to routine application in breeding programs. SSRs or microsatellite markers are proved to be ideal for making genetic maps (Islam, 2004; Niones, 2004), assisting selection (Bhuiyan, 2005) and studying genetic diversity in germplasm. Microsatellite marker analysis is promising to identify major gene locus for

BLB resistance that can be helpful for plant breeders to develop new cultivar.

Nucleotide Diversity Analysis of Three Major Bacterial Blight Resistance Genes in Rice

For combating the disease, the most effective and economical measure is exploitation of host plant resistance. To date, more than 38 R genes for BB resistance have been reported (Chen et al., 2008; Kumar et al., 2012). Diversity analysis of these genes in natural population will facilitate identification of allelic variations which can be exploited in resistance breeding programs (Iyer-Pascuzzi et al., 2007). Recent studies conclude that nucleotide changes in the non-coding and regulatory sites of R genes also contribute to resistance or susceptibility phenotypes of a disease in addition to nucleotide variations in the coding region (Gu et al., 2005; Romer et al., 2009).

Resistance to Bacterial Blight by Suppressing Auxine Biosynthesis in Rice

IAA, the major form of auxin in rice, is generally believed to play an important role in plant growth and development (Teale et al., 2006; McSteen et al., 2010). However, recent studies demonstrate that IAA acts as a negative regulator in the plant immune response (Yang et al., 2013; Navarro et al., 2006), as exogenous application of IAA or auxin analogs in rice and *Arabidopsis* significantly promotes disease symptoms. Treatment with IAA and 2,4-dichlorophenoxyacetic acid (2, 4-D; an analog of IAA) in rice resistant to various types of bacterial blight significantly stimulates phytopathogenic *Xoo* proliferation, resulting in high susceptibility to these compounds (Ding et al., 2008). Similarly, treatment of resistant rice plants with IAA enhances the infectivity of *Xanthomonas oryzae* pv. *oryzicola* (*Xoo*) and *Magnaporthe oryzae* on rice (Fu et al., 2011). In addition, exogenous application of 1-naphthalacetic acid (NAA) or 2,4-D on *Arabidopsis* accelerates the development of disease symptoms during infection by *Pseudomonas syringae* pv. *tomato* (Pto) DC3000 or *Pseudomonas syringae* pv. *maculicola* (Chen et al., 2007; Wang et al., 2007).

Genetic Analysis and Molecular Mapping of QTLs Associated with Resistance to Bacterial Blight in Rice

The International Rice Research Institute has developed a series of near isogenic lines (NILs) which harbor various X genes (IRBB NILs) by using the susceptible cultivar, IR24, as the recurrent parent (Huang et al., 1997). Recently, the molecular markers linking Xa genes in IRBB NILs have been developed using comparative map methods for improving the resistance of commercial cultivars (Kottapalli et al., 2006; Sama et al., 2014). However, climate change has been proposed to affect the microflora of *Xoo* in the field, life cycle, and even the evolution of the pathogen (Garrett et al., 2006; Coakley et al., 1999). Our previous results also revealed that IRBB lines containing *Xa5* or *Xa7* showed moderate resistance, while the NILs harboring a single Xa gene were highly susceptible after the inoculation of a Taiwanese local pathogen, suggesting that more Xa genes are necessary to provide resistance (Wang and Wang 2009). Recently, a durable and broad-spectrum resistance was reported by transmitting one resistance gene and pyramiding with 2–3 other resistance genes (Li et al., 2001; Perumalsamy et al., 2010).

Screening of Rice Varieties for Bacterial Leaf Blight Resistance by Using SSR Markers

Since the bacterial races vary continually influenced by the artificial and natural selection of genes resistance to bacterial blight, it is critical to explore and identify the new resistant resources to control the changeful races (Xia et al., 2012). Since the chemical control is not effective, the utilization of resistant varieties carrying resistance genes have been considered to be the most effective way to control the disease (Nino-Lui et al., 2006). Several molecular markers viz. RFLP, RAPD, SSRs, ISSRs, AFLP and SNPs are presently available to assess the variability and diversity at molecular level (Joshi et al., 2000). With the development of a wide range of molecular techniques, marker assisted breeding is now used to enhance traditional breeding programs to improve crops (Frey et al., 2004). Among various PCR based markers, SSR markers are more popular in rice because they are highly informative, mostly mono locus, co-dominant, easily analyzed and cost effective (Gracia et al., 2004). Simple Sequence Repeats (SSRs) or microsatellites are most suited to routine application in breeding

programs. SSRs or microsatellite markers are proved to be ideal for making genetic maps (Islam, 2004; Niones, 2004), assisting selection (Bhuiyan, 2005) and studying genetic diversity in germplasm. Microsatellite marker analysis is promising to identify major gene locus for BLB resistance that can be helpful for plant breeders to develop new cultivar. Bangladeshi rice varieties have been developed traditionally by selection, hybridization and back crossing with locally adapted high-yielding lines. The conventional methods of plant selection for BLB resistance are not easy because of the large effects of the environment and the low narrow sense heritability of BLB resistance.

Effect of Bioagent Application Time Against Bacterial Leaf Blight of Rice

Besides disease management, bioagents also stimulate plant growth, even if there is no disease, which results in better yield (Mishra and Sinha, 2000). Antagonistic potential of different bioagents against bacterial leaf blight of rice has been reported by several workers (Manmeet and Thind, 2002; Babu and Thind, 2005; Palaniyandi et al., 2006; Gangwar and Sinha, 2012a,b,c and Gangwar, 2013a,b). Time of application of bioagents may have effect on efficacy of bioagents as these are living entity and need a period of time for upsurge optimum population and establish on host. The level of management of disease depends on time of application of bioagents. Influence of time of application of bioagents in plant disease management was studied by several workers including Sindhan et al. (1997) and Vidhyasekaran et al. (2001). Present study was carried out to test the effect of time of application on efficacy of *T. harzianum* and *P. fluorescens* formulations against bacterial leaf blight of rice under field conditions.

REFERENCES

- [1] Babu AGC, Thind BS. 2005. Potential use of combinations of *Pantoea agglomerans*, *Pseudomonas fluorescens* and *Bacillus subtilis* as biocontrol agents for the control of bacterial blight of rice. *Annals of the Sri Lanka, department of agriculture*, 7: 23-37.
- [2] Bhuiyan MAR. 2005. Efficiency in evaluating salt tolerance in rice using phenotypic and marker assisted selection. M. S. Dissertation, Department of Genetics and Plant Breeding, Bangladesh Agricultural University, Mymensingh, Bangladesh. 96p.
- [3] Chen S, Huang ZH, Zeng LX, Yang JY, Liu QG, Zhu X. 2008. High resolution mapping and gene prediction of *Xanthomonas oryzae* pv. *oryzae* resistance gene Xa7. *Mol Breed*. 2008; 22: 433–441.
- [4] Chen Z, Agnew JL, Cohen JD, He P, Shan L, Sheen J, et al., 2007. *Pseudomonas syringae* type III effector AvrRpt2 alters *Arabidopsis thaliana* auxin physiology. *Proc Natl Acad Sci USA*.
- [5] Coakley SM, Scherm H, Chakraborty S. 1999. Climate change and plant disease management. *Annu Rev Phytopathol* 37:399–426.
- [6] Ding XH, Cao YL, Huang LL, Zhao J, Xu CG, Li XH, et al., 2008. Activation of the indole-3-acetic acid–amido synthetase GH3-8 suppresses expansin expression and promotes salicylate- and jasmonate-independent basal immunity in rice. *The Plant Cell*. 20: 228–240.
- [7] Frey JE, Frey B, Sauer C, Kellerhals M. 2004. Efficient low cost DNA extraction and multiplex fluorescent PCR method for marker assisted selection in reeding. *Plant Breed*. 123: 554-557.
- [8] Frey, JE, Frey B, Sauer C, Kellerhals M. 2004. Efficient low cost DNA extraction and multiplex fluorescent PCR method for marker assisted selection in breeding. *Plant Breed*. 123: 554-557
- [9] Fu J, Liu HB, Li Y, Yu HH, Li XH, Xiao JH, et al., 2011. Manipulating broad-spectrum disease resistance by suppressing pathogen-induced auxin accumulation in rice. *Plant Physiol*. 155: 589–602.
- [10] Gangwar GP, Sinha AP. 2012a. Comparative antagonistic potential of fungal and bacterial bioagents against *Xanthomonas oryzae* pv. *oryzae*. *Ann. Pl. Protec. Sci.*,20(1): 154-159.
- [11] Gangwar GP, Sinha AP. 2012b. Evaluation of *Trichoderma* spp. and fluorescent *pseudomonads* for the management of bacterial leaf blight of rice. *Indian Phytopath.*, 65 (1): 89-91.
- [12] Gangwar GP, Sinha AP. 2012c. Effect of time of application on fungal and bacterial bioagents against bacterial leaf blight of rice. *Agric. Sci. Digest.*, 32(2): 123-127.
- [13] Gangwar GP. 2013a. Efficacy of different isolates of fluorescent *pseudomonads* against bacterial leaf blight of rice. *Afr. J. Agric. Res.*, 8(37): 4588-4591.
- [14] Gangwar GP. 2013b. Field efficacy of formulation of fungal bioagents against bacterial leaf blight of rice caused by *Xanthomonas oryzae* pv. *oryzae* (Uyeda and Ishiyama) Dowson. *J. Appl. & Nat. Sci.*, 5(2): 423-426.
- [15] Garrett KA, Dendy SP, Frank EE, Rouse MN, Travers SE. 2006. Climate change effects on plant disease: genomes to ecosystems. *Annu Rev Phytopathol* 44:489–509.
- [16] Gracia AAF, Benchimol LL, Antonica M M, Geraldi IO, Deuza AP. 2004. Comparison of RAPD, RFLP, AFLP and SSR marker for diversity studies in tropical maize inbred lines. *Euphytica* 108: 53-63
- [17] Gracia AAF, Benchimol LL, Antonica MM, Geraldi, IO, Deuza, AP. 2004. Comparison of RAPD, RFLP, AFLP and SSR marker for diversity studies in tropical maize inbred lines. *Euphytica* 108: 53-63.

- [18] Gu K, Yang B, Tian D, Wu L, Wang D, Sreekala C, et al., 2005. R-gene expression induced by a type-III effector triggers disease resistance in rice. *Nature*. 35: 1122–1125. PMID:15973413
- [19] Hsieh SPY. 2003. Rice bacterial blight. In: Cheng CH (ed) *Plant protection illustrations 8: Rice Protection (The next book)*. Bureau of Animal and Plant Health Inspection and Quarantine, Council of Agriculture, Executive Yuan. Taipei, Taiwan, pp 317–338
- [20] Huang CL, Hwang SY, Chiang YC, Lin TP. 2008. Molecular Evolution of the Pi-ta Gene Resistant to Rice Blast in Wild Rice (*Oryza rufipogon*). *Genet*. 179:1527–1538. doi:10.1534/genetics.108.089805 PMID:18622033
- [21] Huang N, Angeles ER, Domingo J, Magpantay G, Singh S, Zhang G, Kumaravadivel N, Bennett J, Khush GS. 1997. Pyramiding of bacterial blight resistance genes in rice: marker-assisted selection using RFLP and PCR. *Theor Appl Genet* 95:313–320
- [22] Islam MM. 2004. Mapping salinity tolerance genes in rice (*Oryza sativa* L.) at reproductive stage. Ph. D. dissertation. University of the Philippines Los Banos, College, Laguna, Philippines. 150 p.
- [23] Islam MM. 2004. Mapping salinity tolerance genes in rice (*Oryza sativa* L.) at reproductive stage. Ph. D. dissertation. University of the Philippines Los Banos, College, Laguna, Philippines. 150 p.
- [24] Johnson R. 1984. A critical analysis of durable resistance. *Annu Rev Phytopathol* 22:309–330
- [25] Joshi SP, Gupta VS, Aggarwal RK, Ranjekar PK, Brar DS. 2000. Genetic diversity and phylogenetic relationship as revealed by Inter simple sequence repeat polymorphism in the genus *Oryza*. *Theor. Appl. Genet.* 100:1311-1320.
- [26] Khush GS, Mackill DJ, Sidhu GS. 1989. Breeding rice for resistance to bacterial blight. *Bacterial Blight of Rice*. IRRI, Manila, pp 207–217
- [27] Kottapalli KR, Sarla N, Kikuchi S. 2006. In silico insight into two rice chromosomal regions associated with submergence tolerance and resistance to bacterial leaf blight and gall midge. *Biotechnol Adv* 24(6):561–589.
- [28] Kumar PN, Sujatha K, Laha GS, Rao KS, Mishra B, Viraktamath BC et al., 2012. Identification and fine-mapping of Xa33, a novel gene for resistance to *Xanthomonas oryzae* pv. *oryzae*. *Phytopathol.* 2012; 102:222–228.
- [29] Li ZK, Sanchez A, Angeles E, Singh S, Domingo J, Huang N, Khush GS. 2001. Are the dominant and recessive plant disease resistance genes similar? A case study of rice R genes and *Xanthomonas oryzae* pv. *oryzae* races. *Genetics* 159(2):757–765
- [30] Manmeet M, Thind BS. 2002. Management of bacterial blight of rice with bioagents. *Plant Dis. Res.*, 17(1): 21-28.
- [31] McSteen P. 2010. Auxin and monocot development. *Cold Spring Harb Perspect Biol.* 2(3): a001479.
- [32] Mew, TW. 1987. Current status and future prospects of research on bacterial blight of rice. *Annu. Rev. Phytopathol.* 25, 359–382.
- [33] Navarro L, Dunoyer P, Jay F, Arnold B, Dharmasiri N, Estelle M, et al., 2006. A plant miRNA contributes to antibacterial resistance by repressing auxin signaling. *Science*. 312: 436–439.
- [34] Nino-Liu DO, Ronald PC, Bogdanove AJ. 2006. *Xanthomonas oryzae* pathovars: model pathogens of a model crop. *Mol. Plant Pathol.* 7, 303–324.
- [35] Nino-Liu DO, Ronald PC, Bogdanove AJ. 2006. Pathogen profile *Xanthomonas oryzae* pathovars: model pathogens of a model crop. *Molec. Plant Pathol.* 7(5): 303-324
- [36] Niones JM. 2004. Fine mapping of the salinity tolerance gene on chromosome 1 of rice (*Oryza sativa* L.) using near-isogenic lines. M.S. dissertation. University of the Philippines Los Banos, College, Laguna, Philippines. 78 p.
- [37] Niones JM. 2004. Fine mapping of the salinity tolerance gene on chromosome 1 of rice (*Oryza sativa* L.) using near-isogenic lines. M.S. dissertation. University of the Philippines Los Banos, College, Laguna, Philippines. 78 p.
- [38] Palaniyandi V, Immanuel JE, Gnanamanickam SS, Thomashow L. 2006. Biological control of rice bacterial blight by plant associated bacteria producing 2,4-diacetylphloroglucinol. *Canadian Journal of Microbiology*, 52(1): 56-65.
- [39] Perumalsamy S, Bharani M, Sudha M, Nagarajan P, Arul L, Saraswathi R, Balasubramanian P, Ramalingam J. 2010. Functional marker-assisted selection for bacterial leaf blight resistance genes in rice (*Oryza sativa* L.). *Plant Breed* 129(4):400–406
- [40] Romer P, Recht S, Lahaye T. 2009. A single plant resistance gene promoter engineered to recognize multiple TAL effectors from disparate pathogens. *Proc Natl Acad Sci USA*. 106: 20526–20531.
- [41] Sama VS, Rawat N, Sundaram RM, Himabindu K, Naik BS, Viraktamath BC, Bentur JS. 2014. A putative candidate for the recessive gall midge resistance gene gm3 in rice identified and validated. *Theor Appl Genet* 127(1):113–124.
- [42] Shen Y, Ronald P. 2002. Molecular determinants of disease and resistance in interactions of *Xanthomonas oryzae* pv. *oryzae* and rice. *Microbes Infect/Inst Pasteur* 4(13):1361–1367
- [43] Sindhan GS, Parasher RD, Hooda I. 1997. Biological control of bacterial leaf blight of rice caused by *Xanthomonas oryzae* pv. *oryzae*. *Plant Dis. Res.*, 12(1): 29-32.
- [44] Teale WD, Paponov IA, Palme K. 2006. Auxin in action: signalling, transport and the control of plant growth and development. *Nature Reviews Mol Cell Biol.* 7: 847–859. PMID:16990790
- [45] Vidhyasekaran P, Kamala N, Ramanathan P, Rajappan K, Paranidharan V, Velazhahan R. 2001. Induction of systemic resistance by *Pseudomonas fluorescens* Pf1 against *Xanthomonas oryzae* pv. *oryzae* in rice leaves. *Phytoparasitica*. 29(2): 155-166.
- [46] Wang CS, Wang AZ, Lin DG. 2013. The application of mutants in breeding disease resistance in rice. Paper presented at the Special issue or the symposium on important crop pathogen detection and management, Taichung
- [47] Wang D, Pajerowska-Mukhtar K, Culler AH, Dong X. 2007. Salicylic acid inhibits pathogen growth in plants through repression of the auxin signaling pathway. *Curr Biol.* 17: 1784–1790.
- [48] Xia C, Chen, H, Zhu X. 2012. Identification, Mapping, Isolation of the Genes Resisting to Bacterial Blight and Breeding Application in Rice. *Molecular Plant Breeding*, 3(12): 120-130.
- [49] Yang DL, Yang YN, He ZH. 2013. Roles of plant hormones and their interplay in rice immunity. *Mol Plant.* 6: 675–685.
- [50] Yang Z, Sun X, Wang S, Zhang Q. 2003. Genetic and physical mapping of a new gene for bacterial blight resistance in rice. *Theor Appl Genet* 106(8):1467–1472