

National Conference on Recent Trends in Synthesis and Characterization of Futuristic Material in Science for the Development of Society (NCRDAMDS-2018)

In association with International Journal of Scientific Research in Science and Technology



Isolation of Fungi From Root Tubers Of Chlorophytum Borivilianum San. And Fem.

Anil N. Korpenwar¹, Deepmala A .Gaikwad²

¹Rashtrapita Mahatma Gandhi Science and Arts College, Nagbhid, Chandrapur, Maharashtra, India ²Shri Shivaji Science and Arts College Chikhli, Buldana, Maharashtra, India

ABSTRACT

Chlorophytum borivilianum root tubers are used as a raw material for the preparation of some important drugs for curing various human diseases. During unscientific methods of storage causing fungal contamination .The fungal contamination affect on the chemical composition of raw materials and thereby decreases potency of drugs. Regarding the above fact the present experiment was conducted and observed that the maximum fifteen fungi were isolated by using Agar Plate Method. And minimum thirteen fungi were isolated by using Blotter Method respectively.

Keywords: Chlorophytum borivilianum, Isolation, Fungal Contamination.

I. INTRODUCTION

Chlorophytum borivilianum (San .and Fem) is commonly known as safed musli, family liliaceae, is a highly economical important medicinal plant in Indian system of Ayurveda. (Chakraborthy et al., 2014). (Purohit and Prajapati, 2003) reported that in Indian herbal medicine used as an aphrodisiac, antiageing health restorative and health promoter. Varying its common use for health promotion, it is also used for increasing lactation, treating various gynecological disorders, arthritic conditions and to control diabetes mellitus.. Its medicinally important root tubers are rich alkaloids, many vitamins, minerals, source of proteins ,carbohydrates, saponins and steroids (Gondi et al., 2004). World Health Organization (WHO) survey report indicates that about 70-80% of the world population particularly in the developing countries rely on non-conventional medicines mainly of herbal sources in their primary health (Okunlola et al., 2007). The herbal medicines are prepared from materials of plant origin they are prone to microbial contamination (Shrikumar and Ravi, 2007). The presence of microbial contaminant in non sterile pharmaceutical products can reduce the therapeutic activity of the products and adversely affect on patients taking the medicines.

Manipulation and processing factors largely indicate the microbiological quality of the products (Busse ,2000).

Many researchers have confirmed that the presence of potential contaminants in herbal preparations viz.(Czech et al., 2001), (Kulshrestha et al .,2008), (Kosalec et al., 2009),(Martins et al., 2001), (Alwakeel ,2008), (Idu et al., 2011). Thus, (Okunlola et al., 2007) concluded that the manufacturers should ensure the lowest possible level of microorganisms in the raw material, finished dosage forms and the packaging components to maintain appropriate quality, safety and efficacy of the natural products.(Rai and Mehrotra, 2005) studied that the quality of herbal drugs with lots of contaminants like heavy metals, pesticides and microbes causes various deformities like congenital paralysis, sensori-neural defects, liver and kidney damage .The unscientific methods of harvesting, collection, storage of raw materials, post harvest processing, transport and storage of herbal drugs in unhygienic conditions, are the main causes considered to make both, raw materials as well as herbal drugs prone to microbial infections leading to deterioration. It can cause health hazard. So present investigation is an attempt to identify the mycofloa associated with Chlorophytum borivilianum root tubers.

II. MATERIALS AND METHODS

1. Collection of plant material.

Chlorophytum borivilianum root tubers were collected from different authentic stores of Jalna district in presterilized polythene bags and brought to the laboratory. samples were identified using the Flora of Marathwada (Naik, 1998) at Department of Botany, Dr. Babasaheb Ambedkar Marathwada University .The plant material was first cleaned by washing several times under running tap water and Surface sterilization was performed by sequentially rinsing the plant material with 70% ethanol for 30 seconds, then with 0.01% mercuric chloride for 5 minutes and finally with sterile distilled water for 2-3 times, then dried in between folds of sterile filter papers, placed at equal distance on moist blotters on the sterilized petriplates similarly material inoculated aseptically on the sterilized petriplates containing Potato Dextrose Agar (PDA) medium and Blotter Method incubated at25±2°C temperature for 7 days.

2. Isolation of mycoflora.

Mycoflora was isolated by using Blotter Method and Potato Dextrose Agar (PDA) medium .

3. Identification of fungi

The fungi occurring on plant material in the plates were identified preliminary on the basis of sporulation characters like sexual or asexual spores with the help of stereoscopic binocular microscope. The identification and further confirmation of fungi was made by preparing slides of the fungal growth and observing them under compound microscope. The identification was made with the help of manuals (Mukadam et al., 2006) , (Alexopoulous, 1996) and (Barnett,1970) Pure cultures of these fungi were prepared and maintained on potato dextrose agar (PDA) slants.

III. RESULTS AND DISCUSSION

1. Incidence of fungi on tuber roots of Chlorophytum borivilianum:

In order to study the incidence of fungi on tuber roots of Chlorophytum borivilianum was studied and the results are summarized in table 1.

From the result given in table 1, it is indicated that young root tubers of Chlorophytum borivilianum showed no incidence of fungi except Aspergillus niger and Fusarium oxysporum on blotter method where as the same young root tubers kept on PDA showed slight high incidence of fungi. In case of mature root tubers Chlorophytum borivilianum the blotter paper method showed the incidence of fungi like A. flavus, A. niger and f. oxysporum where as PDA method showed slight higher incidence of fungi as compared blotter method.

Table 1. Incidence of fungi on root tubers of
Chlorophytum borivilianum from different age.

	Vouna					
Eunci	Young		Mature		Stored	
Fungi	Blotte	PD	Blott	PD	Blott	PD
	r	Α	er	Α	er	Α
Alternaria	-	05	-	-	10	10
alternate						
Aspergillu	-	-	10	10	20	35
s flavus						
Aspergillu	05	15	15	35	25	50
s niger						
Aspergillu		07		20	05	20
S	-	05	-	20	05	30
fumigates						
Aspergillu						05
S	-	-	-	-	-	05
nidulance						
Aspergillu	-	-	-	-	10	15
s terreus						
Curvularia	-	10	-	20	05	25
lunata						
Cladospori	-	-	-	-	-	20
um.sp.						
Fusarium	05	15	15	25	25	45
oxysporum						
Fusarium	-	10	-	15	10	15
roseum						
Mucor	-	-	-	-	05	10
globsus					05	
Phoma sp.	-	-	-	-	05	15
Penicilliu	-		-		05	30
m notatum		-		-	05	30
Rhizopus	_	10	-	20	10	35
stolonifer		10		20	10	55
Rhizoctoni	_		-		05	05
a solani		-		-		05

It was interesting to note that the stored root tubers of Chlorophytum borivilianum, the maximum incidence of fungi were reported as compared to young and mature root tubers. In case of stored fruits PDA method , percent incidence of fifteen fungi such as Alternaria alternata (10) Aspergillus flavus (35), Aspergillus niger (50), Aspergillus fumigates (30), Aspergillus nidulance (05), Aspergillus terreus(15), Curvularia lunata (25), Cladosporium.sp (20), Fusarium oxysporum (45), Fusarium roseum(15), Mucor globsus (10), Phoma sp.(15), Penicillium notatum (30), Rhizopus stolonifer (35) and Rhizoctonia solani (05) were reported where as thirteen fungi viz. Alternaria alternata (10) Aspergillus flavus (20), Aspergillus niger (25), Aspergillus fumigates (05), Aspergillus terreus(10), Curvularia lunata (05), Fusarium oxysporum (25), Fusarium roseum(10), Mucor globsus (05), Phoma sp.(05), Penicillium notatum (05), Rhizopus stolonifer (10), Rhizoctonia solani (05) were associated on Blotter method.

Roy, (2003) reported that the frequent occurrence of Aspergillus, Fusarium and Penicillium species on different crude herbal drugs. (Abou et al., 1999) and (Gautam and Bhadauria, 2008, 2010) noted that the occurrence of these fungi in Egyptian herbal drugs and in stored herbal fruit samples.(Dhale, 2013)studied and concluded that 45 fungi were recorded on the blotter and agar plate methods . Maximum number of fungi belonged to Deutroomycetes. All samples of plant material showed maximum infestation of A. niger and Aspergillus spp. (Sharma et al., 2013) The some herbs are good substrate for Aspergillus flavus infestation and production of aflatoxins with potential hazard to the health of consumers. (Kumar et al., 2009) concluded that the herbal preparations had the presence of fungal contaminants with predominance of Aspergillus spp. and Penicillium spp., but Mucor spp., Candida spp., Trichosporium spp., also were found. The fungal deterioration adversely affects the chemical composition of the raw materials and thereby decreases the medicinal potency ofherbal drugs .respectively, supporting findings of present investigations.

IV. CONCLUSION

The present study was aimed to isolate the mycoflora [1]. associated with root tubers of chlorophytum borivilianum. In the stored root tubers of Chlorophytum borivilianum, the maximum incidence of fungi was reported as compared to young and mature root tubers. This study stresses the importance of scientific methods [2]. for proper storage of plant parts. Therefore, this study suggests that the methods of harvesting, collection, preparing and storage of medicinal plants must be improved for reducing percentage incidence of [3]. mycoflora and mycotoxins contaminations. These

factors can certainly contribute significantly promoting ecofriendly herbal drugs for the health care of human society.

Photo plate 1. Isolation of Mycoflora on different media.



Blotter Method.



PDA Medium.

V. ACKNOWLEDGEMENT

The authors are thankful to principal, Shri Shivaji Science and Arts College ,Chikhli. for providing the necessary laboratory facilities.

VI. REFERENCES

- Abou, A.A., Soliman, K. M., El-Tantaway, M. E., Ismail, B. R.and Naguib, K.(1999).Quantity estimation of some contaminants in commonly used medicinal plants in Egyptian market. Food Chemistry ,67 : 357-363.
- Alexopolous, C. J. (1996). Introductory To Mycology, John Wiley and Sons, Inc.Publication, New York Winchester, Brisbane ,Tornoto And Singapore.
 - Alwakeel, S. S. (2008). Microbial and heavy metals contamination of herbal medicines.

Research Journal of Microbiology, 3(12): 683-691.

- [4]. Barnett, H.C.(1970).Illustrated genera of Fungi. [16]. imPerfecti, Burges Publication, Minn, USA.
- [5]. Busse, W. (2000). The significance of quality for efficacy and safety of herbal medicinal products. Drug Information Journal, 34: 15-23.
- [6]. Chakraborthy G.S., Vidhu Aeri., Pawan [17]. Vermaand., Sarita Singh. (2014) phytochemical and antimicrobial studies of chlorophytum borivilianum .Pharmacophore, Vol. 5 (2):258- [18]. 261.
- [7]. Czech, E., Kneifel, W., and Kopp, B. (2001). [19]. Microbiological status of commercially available medicinal herbal drugs- A screening study. Planta Medica, 67: 263-269.
- [8]. Dhale ,D. A. (2013) surface mycoflora of stored part of herbal medicine. Int J pharm Bio Sci ,4(3): [20].
 (B) 568-574.
- [9]. Gautam A. K. and Bhadauria, R. (2008).
 Occurrence oftoxigenic moulds and mycotoxins in [21]. ayurvedic medicine Trifla churn. J. Indian Mycol. Pathol, 38 :664-666.
- [10]. Gautam, A. K., and Bhadauria, R. (2010). Fungal [22]. andmycotoxin contamination of some common stored herbal fruit samples. J. Indian Bot. Soc,89 : 74-79. [23].
- [11]. Gondi, M., Tyagi, S.K., and Srinivasan, K. (2004). In Safed Musli. A white Gold. Agrobios (India) Publ.House, Jodhpur: 11-19.
- [12]. Idu, M., Erhabor, J. O., and Idele, S. O. (2011). Microbial load of some medicinal plants sold in local markets of Benin City, Nigeria. International [24]. Journal of Medicinal and Aromatic Plants, 1(3): 272-277.
- [13]. Kosalec, I., Cvek, J., and Tomic, S. (2009). Contaminants of medicinal herbs and herbal products. Archives of Industrial Hygiene and Toxicology, 60:485-501.
- [14]. Kulshrestha, R., Gupta, C. P., Shukla, G., Kundu, M. G., Bhatnagar, S. P., and Katiyar, C. K. (2008). The effect of water activity and storage temperature on the growth of Aspergillus flavus in medicinal herbs. Planta Medica, 74: 1308-1315.
- [15]. Kumar, A., Shukla, R., Singh, P., and Dubey, N. K. (2009). Biodeterioration of some herbal raw materials by storage fungi and aflatoxin and assessment of ymbopogon flexuosus essential oil

and its components as antifungal. International Biodeterioration &Biodegradation, 63:712-716.

- Martins, H. M., Martins, M. L., Dias, M. I., & Bernardo, F. (2001). Evaluation of microbiological quality of medicinal plants used in natural infusions. International Journal of Food Microbiology, 68: 149-153.
- Mukadam ,D.S., Patil, M.S., Chavan, A.M., Patil, A.R. (2006) The Illustrations Of Fungi.Sarraswati Printing Press.Aurangabad,1-254.
- Naik, V.N. (1998).Flora of Marathwad, Amrut Prakashan, Aurangabad.(M.S) India.
- Okunlola, A., Adewoyin, B. A., and Odeku, A. O. (2007). Evaluation of pharmaceutical and microbial qualities of some herbal medicinal products in South Western Nigeria. Tropical. Journal of Pharmaceutical Research, 6: 661-670.
- 20]. Purohit, S.S and Prajapati, N.D. (2003), Agro's Colour Atlas of Medicinal Plants, Agrobios Publications, Jodhpur, 43.
- [21]. Rai ,V. and Mehrotra, S. (2005) Toxic contaminants in herbal drugs.Enviro. News Archives, 11(4): 1-3.
- [22]. Roy, A. K. (2003). Mycological problems of crude herbal drugs: Overviewand challenges. Indian Phytopath ,56: 1-13.
- [23]. Sharma Sumedha ,Dimple Gupta and Y.P. Sharma. (2013) Aflatoxin Contamination In Chilgoza Pine Nuts (Pinus gerardiana Wall.) Commercially Available In Retail Markets of Jammu. India. Int J Pharm Bio Sci Apr, 4(2): (B) 751-759.
- Shrikumar, S. and Ravi, T.K. (2007) Approaches towards development and promotion of herbal drugs. Pharmacognosy Reviews,1(1):180-183.