

# Influence of the Topical Application Acetone Solution of Vitamin A (Retinol) to the Fifth Instar Larvae of the Silkworm, *Bombyx mori* (L) (Race : PM x CSR2) on the Economic Parameters

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# ABSTRACT

The attempt concerned with efficient topical application of acetone solution of Vitamin A (retinol) to the fifth instar larvae of multivoltine silkworm, *Bombyx mori* (L) (Race: PM x CSR2) and assessment of economic parameters. The experimentatio was carried at laboratory level. The ten microliters of one milligram per liter strength acetone solution of retinol was topically applied to the individual fifth instar larvae of multivoltine cross breed race of silkworm, *Bombyx mori* (L) (PM x CSR2) variously at 48, 54, 60 and 66 hours after the fourth moult. Topical application of acetone solution of retinol was found variously reflected into prolongation of larval age (14 - 39 %); improvement in the tissue somatic index (TSI) of silk glands (3.06 - 3.343); cocoon shell ratio (2.790 - 3.177) and denier scale of silk filament (p < 0.01). Topical application of acetone solution of retinol, thus chiefly lengthen the larval age in silkworm, *Bombyx mori* (L). Time required for eating and amount of mulberry leaves eaten had both been increased and they were practically reflected into the improvement in the quality of cocoon and the silk filament. Diterpenoid nature of retinol and delay in the spinning among the larvae, recipient of retinol in the attempt indicate the possibility of insect juvenoid activity of retinol in silkworm, *Bombyx mori* (L). Efficient utilization of retinol, the diterpenoid through acetone solvent for topical application to the fifth instared larvae of silkworm, *Bombyx mori* (L) may open a new biotechnological avenue in the sericulture industries.

Keywords : Retinol, TSI, Shell Ratio, Denier Scale of Silk Filament.

# I. INTRODUCTION

Vitamins are the organic compounds required by organism a a vital nutrient in limited amounts. Supplementation of vitamins serve to orchestrate the metabolism. The larvae of silkworm, Bombyx mori (L) deserve appreciation for synthesis of silk for it's metamorphosis. Sericultural practices are serving a lot to provide the silk fibre. The silkworm, *Bombyx mori* (L) exert a significant influence on the concept of insect metamorphosis through it's simple life cycle and efficient utilization of the nutrients from the mulberry, *Morus alba* (L). Interplay of juvenile hormone and moulting

hormone in the insect larval body serves to orchestrate the progression of metamorphosis from one instar to next, with moulting hormone regulating the onset and timing of moulting cycle and juvenile hormone regulating the quality of moult (Riddiford, 1985, 1994; Sehnal, 1985). During the last larval stadium of holometabolous insects, such as silkworm, *Bombyx mori* (L), a reduction of JH in haemolymph is the necessary step in the initiation of metamorphosis. It has been demonstrated that, haemolymph ecdysteroid and JH level undergo the developmental changes during larval - larval and larval - pupal cycles in silkworm, *Bombyx mori* (L) (Calvez <u>et al</u>, 1976; Kiguchi and Agui, 1981 ). Juvenoids are well known in prolonging the larval life in the insect and have long been tried for qualitative improvement of silk (Ratnasen, 1988; Granier and Granier, 1983 and Mamatha et al, 1999 ). There is considerable evidence that juvenile hormone mimics occur in plants, which occasionally leads to economically important consequences in the insect development 1979 ). Juvenile hormone active (Slama. compounds are found in many higher plants, exogenous application through suitable solvents of which exhibited potent activity in the insects (Prabhu and John, 1975). Efficient use of available system, the principle of quality improvement made man to use juvenoids for pest control as well as for the silk yield. Use of juvenoids (synthetic, plant derived and animal derived ) in the rearing of silkworm larvae had positive influence, especially in the silk yield (Ching et al, 1972; Nihmura et al, 1972; Muroga et al, 1975; Kamada and Shimada, 1988 ; Rajashekhargouda, 1991 ; Vitthalrao et al, 2002, 2003 and Vitthalrao, 2004)

Retinoic acid and insect juvenile hormone (JH) are structurally related terpenoids, which are widespread in nature and are involved in much more biological activities including morphogenesis, embryogenesis and cellular differentiation. The retinol is a diterpenoid, a terpenoid derived from a diterpene, which include the compounds with C20 skeleton of the parent diterpene, which has been rearranged or modified by the removal of one or more skeletal atoms (generally, methyl groups) ( The retinoids deserve important role in the process of morphogenesis and in immune response in the insects like R. prolixus, suggesting that the molecular mechanism recognize the terpenoid backbone as one of the important structural determinant in insects CHEBI: ( 23849 http://www.ebi.ac.uk/chebi/searchId.do?chebiId=C HEBI:23849) Angelica Nakamura, et al, 2007). Terpenoid hormones seems to act as the morphogens throughout the The metazoan.

regulatory activities of terpenoid hormones range from controlling metamorphosis in insects (Riddiford and Ashburner, 1990) and to determine the germ cell fate in the mammalians (Bowels, et al , 2006). In the metamorphosis, the interplay of the juvenile hormone and ecdysone serve to orchestrate the progression from one instar to the next, with ecdyosteroid regulating the onset and timing of the moult and JH determining whether the moult would be larval - larval or larval - pupal (Gilbert, et al, 1996; Mamatha, et al, 2005). Phytophagous insects like silkworm, Bombyx mori (L) derive their juvenoid nutrients through the plant material available for them (Khyade, et al , 2007). Retinol like vitamin nutrients may either be synthesized by the insect tissue or derived from the plant material. Nutrition with vitamins is playing important role in the improvement of growth and development in silkworm, Bombyx mori (L).

Juvenoids are known for disruption of normal developmental pattern leading to the deformities in the insects. Interestingly, the silkworm, Bombyx mori (L) is known to have a stimulatory influence on the administration of exogenous Juvenoids (JHA) in a appropriate quantities. The specific titer of juvenoids, either topical or through the food, at the specific period of the larval instars of silkworm, Bombyx mori (L) are positively reflected into the retention of larval features long enough enabling the larvae to consume maximum quantity of mulberry leaves and to synthesize paramount silk to be used in spinning the qualitative cocoon (Akai, et al, 1990; Mamatha, et al, 2005, 2006, 2008; Chowdhary, et al, 1990; Miranda, et al, 2002; Mamatha, et al, 2006, 2008). Diterpene structure, insect juvenoid activity and vitamin nature of Retinol made to plan for the efforts on it's topical application through the acetone to the fifth instar larvae of silkworm, Bombyx mori (L) (PM x CSR2). The attempt was carried at the laboratory level.

#### **II. METHODS AND MATERIAL**

All the facilities at the Sericulture Farm of Agriculture Development Trust, Baramati were utilized for the experimentations. The entire attempt was divided into the parts, which include: Rearing of the silkworm larvae; Topical application of retinol through the acetone solution to the fifth instar larvae; Analysis of parameters (Larval, cocoon and silk filament) and Statistical analysis of the data. The larvae of silkworm belongs to polyvoltine cross breed ( PM  $x \ CSR_2$  ) race were reared in the laboratory through standard methods (Krishnaswami et al, 1978). The vitamin A, retinol was procured from through dealer. One mg of retinol was dissolved in the acetone and the stock solution of one ppm was prepared. Soon after fourth moult, the fifth instared larvae were grouped into one control group; one acetone treated group and four experimental groups, each with hundred individuals. Ten microliters of acetone solution of retinol was used for topical application to the individual larva in each group separately (Table -1). The experimental group: "A" received the topical application of retinol at 48 hours after the fourth moult. The experimental group: "B" received the topical application of retinol at 48 and 54 hours after the fourth moult. The experimental group: "C" received the topical application of retinol at 48; 54 and 60 hours after the fourth moult. And the experimental group: "D" received the topical application of retinol at 48; 54; 60 and 66 hours after the fourth moult. The tender mulberry leaves were soaked separately in each stock solution of digoxin ( at the rate of 100 gm leaves in 400 ml stock solution ) for half an hour. Topical application was followed by feeding the larvae with tender mulberry leaves.

The schedule of feeding was 100 gms of mulberry leaves for each time (48, 54, 60, 66 hours after the fourth moult) for each group of 100 larvae. Acetone treated control and untreated control groups of larvae were also maintained.

Daily larval weight was recorded. For the purpose to calculate tissue somatic index (TSI) of silk glands, ten larvae from each group were selected at random on the fifth day, anesthetized, dissected and silk glands were separated. The silk glands were bottled and weighed on electronic balance. The weight of silk glands was divided by weight of larva. The quotient thus obtained was multiplied by 100. Weight of silk glands and larval body weight, thus, were accounted for the calculation of tissue somatic index (TSI) of silk glands.

The matured larvae ( having transparent skin, feeding stopped and moving its head in specific manner for searching the surface for attachment of fluid silk ) were transferred to the mountage for spinning the cocoon. The larval duration ( right from zero hour of fifth instar to fifty percent spinning ) was recorded. The cocoons were harvested on sixth day after mounting the mature larvae on the mountage. Cocoon weight, shell weight and pupal weight were recorded. Shell ratio was calculated. Ten cocoons per replication were reeled and length (m) of unbroken silk filament was obtained by using eprouvate. Weight of silk filament from individual cocoon was recorded. Length (m) and weight (gm) of silk filament were accounted for calculation of Denier The the scale. experimentation was repeated for thrice for the purpose of consistency in the results. The statistical methods were employed to calculate

the mean, standard deviation, percent variation and student "t" - test ( Norman and Bailey, 1955). The data collected belongs to three successive trials.

# **III. RESULTS AND DISCUSSION**

The larval parameters get reflects on the quality of the cocoon and silk fibre. Extension of fifth instar larval period was observed in groups of larvae topically applied with one ppm acetone solution of retinol. The extension of fifth instar larval period was 14 - 39 % in all the treated groups of larvae. Maximum increase in the larval duration was recorded in larvae received four times (at 48, 54, 60 and 66 hours after the fourth the topical application of moult) retinol. Corresponding to the extension in the larval duration, an increase in the larval growth by the body weight (17-24%) was observed in all the treated groups. Tissue somatic index (TSI) signify the percentage of tissue in entire body. Tissue somatic index (TSI) of the silk glands of treated larvae in the percent study was found significantly increased. Treating the larvae with one ppm retinol solution through acetone at 48, 54, 60 and 66 hours after fourth moult was found variously reflected into most significant improvement in the TSI of silk glands (table 2).

The economic parameter in sericulture is the Cocoon spinned by the mature fifth instar larvae of silkworm, *Bombyx mori* (L). Cocoon is the most important aspect in sericulture as it is used for reeling the commercial silk fibre. Cocoon weight, shell weight and thereby the shell ratio were found influenced by the topical application of acetone solution of retinol to the fifth instar larvae of silkworm, *Bombyx mori* (L). The range of percent increase in the cocoon weight and shell weight in the experimental (treated) groups was 42.911 to 50.268 and 66.565 to 77.243 respectively. Shell ratio of the cocoons was found improved in the corresponding groups of treatment. Most significant ( p < 0.001 ) shell ratio belonged to cocoons harvested from the group of larvae treated with one ppm acetone solution of retinol at 48 hours after the fourth moult. Silk filament is sole aim in sericulture. Length and weight of entire silk filament are the qualitative measurements to be accounted for the Tex and Denier scale. Both the parameters ( Tex and Denier ) of silk filament were found influenced through treating the larvae with retinol solution ( one ppm ). The retinol through acetone was found resulted into fortified silk filament, with reference to Tex and Denier scale. The silk reeled from the cocoons belong to the group C and D ( Table 4 ) were exhibited most significant improvement.

Prolonged larval duration in the larvae fed with digoxin treated mulberry leaves in the present study is as good as tendency of larvae retaining their larval stage. Extension of larval duration is one of the distinguishing features of insect larvae recipient of exogenous juvenoid( Akai and Kobayashi, 1971 ). The larvae of all the treated groups in the present study were found increased in their body weight. The retinol received by larvae through the acetone topically, may influence the appetite, nutrition and absorption of digested food. This may be responsible for accelerated growth of silk glands. Cocoon is the material used for reeling the commercial silk fibre. It is in fact, a protective shell made up of a continuous and long proteinaceous silk filament spun by mature silkworm prior to pupation for self protection from adverse climatic situations and natural enemies. The juvenoid titre ( endogenous and / or exogenous ) in the body of larvae stimulate hypermetabolism (Slama, 1971 ). Use of retinol through the acetone for topical application, thus chiefly reflected into lengthening fifth instar larval duration. The time required for eating and amount of mulberry leaves eaten has both been increased and were practically reflected into the improvement of cocoon quality, shell ratio and silk filament quality. Retinol topically applied may be utilized by the silkworm larvae for the extra synthesis of silk. The retinol is one of the most popular vitamin suppliment used by man. Use of retinol through

acetone for rearing of silkworm larvae is much more new avenue in sericulture for the qualitative cocoon easy method. Use of vitamins like retinol may open a and silk filament.

**Table 1 :** Schedule of topical application of acetone solution ( ten microliteres of one ppm ) of retinol to the fifth instar larvae of silkworm, *Bombyx mori* (L).

Hours after IV	48	54	60	66
moult→				
group↓				
Control	-	-	-	-
А	+	-	-	-
В	+	+	-	-
С	+	+	+	-
D	+	+	+	+

+ indicates topical application of ten microliters of one ppm solution of acetone to individual larvae of silkworm, *Bombyx mori* (L).

- Indicates No Treatment.

$Groups \rightarrow$	Control	А	В	С	D
Parameter					
$\downarrow$					
Larval	156	178**	193**	198**	217**
duration	(±23.786)	(±5.943)	(±4.614)	(±5.651)	(±5.503)
(hours)		14.102	23.717	26.923	39.102
Larval	3.126**	3.658**	3.789**	3.881**	3.781**
weight	(±0.457)	(±0.539)	(±0.438)	(±0.769)	(±0.947)
(gm)		17.018	21.209	24.152	20.953
Weight of	0.748	0.987**	1.014**	1.058**	1.035**
silk glands	(±0.019)	(±0.112)	(±0.236)	(±0.339)	(±0.376)
		31.951	35.561	41.443	38.368
Tissue	23.928	26.981**	26.761**	27.261**	26.373**
somatic		3.061	2.833	3.333	2.445
index (TSI)					

- Each figure is the mean of the three replications.
- Figure with ± sign in the bracket is standard deviation.
- Figure below the standard deviation is the increase for calculated parameter and percent increase for the others over the control.
- \* P < 0.05
- \*\* P < 0.005
- \*\*\* P < 0.01

Table 3 : effect of Retinol on the cocoon parameters of silkworm, *Bombyx mori* (L) (Race : PM x CSR2).

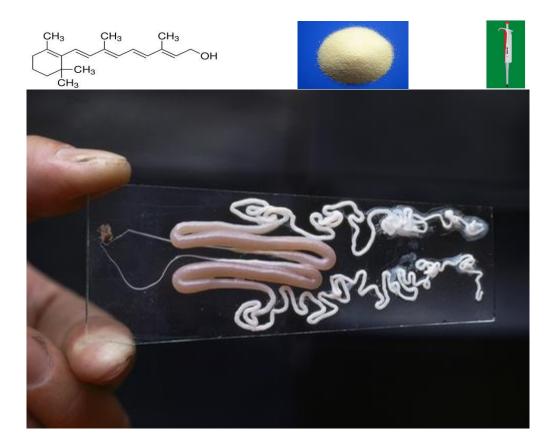
groups→	Control	А	В	С	D
parameters↓					
Cocoon	1.856**	2.671**	2.694**	2.716**	2.789**
weight (gm)	(±0.273)	(±0.445)	(±0.786)	(±0.643)	(±0.671)
		43.911	45.150	46.336	50.269
Shell weight	0.329	0.548***	0.553**	0.571*	0.583*
(mg)	(±0.078)	(±0.102)	(±0.181)	(±0.187)	(±0.105)
		66.565	68.085	73.556	77.203
Pupal weight	1.527	2.123**	2.141**	2.145*	2.206*
(mg)	(±0.093)	(±0.864)	(±0.786)	(±0.654)	(±0.467)
		39.030	40.209	40.471	44.466
Shell ratio	17.726	20.516***	20.527**	21.023*	20.903*
		2.790	2.801	3.297	3.177

- Each figure is the mean of the three replications.
- Figure with ± sign in the bracket is standard deviation.
- Figure below the standard deviation is the increase for calculated parameter and percent increase for the others over the control.
- \* P < 0.05
- \*\* P < 0.005
- \*\*\* P < 0.01

Table 4: Effect of Retinol on the silk filament parameters in silkworm, Bombyx mori (L) (Race : PM x CSR<sub>2</sub>)

group→ parameters↓	Control	А	В	С	D
S.F length (m)	794	958**	963*	959*	976*
	(±23.245)	(±8.679)	(±11.748)	(±11.764)	(±23.789)
		20.654	21.284	20.780	22.921
S.F weight	0.181*	0.254**	0.277*	0.321*	0.324*
(mg)	(0.039)	(±0.083)	(±0.041)	(±0.052)	(±0.086)
		38.043	53.038	77.348	79.005
Tex	0.2273	0.2658*	0.2784*	0.3093*	0.2447**
		0.0385	0.0511	0.082	0.0204
Denier	2.051	2.386**	2.588*	3.012***	2.987***
		0.335	0.537	0.961	0.936

- Each figure is the mean of the three replications.
- Figure with ± sign in the bracket is standard deviation.
- Figure below the standard deviation is the increase for calculated parameter and percent increase for the others over the control.
- \* P < 0.05
- \*\* P < 0.005
- \*\*\* P < 0.01



# **IV. Acknowledgement**

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# **V. CONCLUSION**

The proposed payment system combines the Iris recognition with the visual cryptography by which customer data privacy can be obtained and prevents theft through phishing attack [8]. This method provides best for legitimate user identification. This method can also be implemented in computers using external iris recognition devices.

#### VI. REFERENCES

 Angelica Nakamura; Renata Stiebler; Marcelo R Fantappié; Eliane Fialho; Hatisaburo Masuda and Marcus F Oliveira (2007). Effects of retinoids and juvenoids on moult and on phenoloxidase activity in the blood-sucking insect, Rhodnius prolixus (L). Acta Trop. 2007 Jul 1;: 17686447 Cit:1 http://lib.bioinfo.pl/paper:17686447.

- [2]. Riddiford, L. M. and Ashburner, M. (1990). Role of Juvenile hormone in larval development and metamorphosis in Drosophila melanogaster (L). Comparative Endocrinology. 82: 172-183.
- [3]. Bowles, J.; Knight, D.; Wihlem, D.; Richman, J.; Mamiya, S.; Yashiro, K.; Chawengsaksphak, K.; Wilson, M. J.; Rossant, J.; Hamada, H. and Koopman, P. (2006). Retinoid signaling determines germ cell fate in mice. Science 312: 596 - 600.
- [4]. Ying, H. Wang ; Guirong Wang and Gerold A. LeBlanc (2006). Cloning and characterization of the retinoid X receptor from a primitive crustacean, Daphnia magna (L). General and Comparative Endocrinology. 150: 309 - 318. www.sciencedirect.com and www.elsevier.com/Locate/ygcen
- [5]. Akai, H. and Kobayashi, M. (1971). Introduction of prolonged larval instar by juvenile hormone in Bombyxmori(L) ( Lepidoptera : bombycidae ). Appl. Entemol. Zool, 6: 1938 - 1939.
- [6]. Calvez, B.; Hiren, M. and Reddy, M. (1976). Progress of developmental programme during the last larval instar Bombyxmori (L): Relationships with food intake, ecdysteroids and juvenile hormone. Journal of insect physiology,24 (4): 233-239.

- [7]. Ching, F.C; Mukakeshi, S. and Tamura,S. (1972). Giant cocoon formation in the silkworm, Bombyxmori (L) treated with methyloneoxyphenyl derivatives. Agar. Biol. Chem. 36: 692-694.
- [8]. Granier and Granier (1938). Fenoxycarb, a fairly new growth regulator: A review of its effects on insects. Ann. Appl. Biol. 122: 369-403.
- [9]. Kamada, A. and Shimada, S. (1988). Effects of methoprene on growth profile of larval organs of silkworm, Bombyxmori (L). Jornal of sericulture science, Jpn. 48 (2): 129-136.
- [10]. Khyade V.B. (2004). Influence of juvenoids on silkworm, Bombyxmori (L). Ph.D thesis, Shivaji University, Kolhapur. (Maharashtra, India).
- [11]. Kiguchi, K. and Augi, N. (1981). Ecdysteroid level and development events during larval moulting in silkworm, Bombyxmori (L). insect physiol. 26: 805-813.
- [12]. Krishnaswami, S.; Narimhan, N.; Suryanarayana, S.K and Kumararaja, S. (1978). Manual on sericulture, vol. 2: silkworm rearing, FAO publication, agricultural services bulletin no. 15.
- [13]. Mamatha, D.N; Nagalalakhamma, K.; Vijay, P. and RajeshwaraRao, A.M. (1999). Impact of selected juvenile hormone mimics on organic constituents of silkworm, Bombyxmori (L). proceedings of NSTS- 99: 185-186.
- [14]. Muroga, A.; Nakajima, M.; Aomori,S.; Ozawa, Y. and Nihmura, M. (1975). Utilization of synthetic juvenile hormone analogue for the silkworm rearing on the mulberry leaves. J. Sreic. Sci. Jpn. 44: 267-273.
- [15]. Nihmura, M.; Aomori,S.; Mori, K. and Matusui, M. (1972). Utilization of synthetic compounds with juvenile hormone activity for silkworm rearing. Agri. Biol. Chem. 36: 882-889.
- [16]. Normn, T.J and Bailey, (1995). Stastical methods in biology.
- [17]. Prabhu, V.K.K; John, M. (1975). Juvenomimetic activity in some plants. Experimential, 31: 913-914.
- [18]. Rajashekhargouda, R. (1991). Studies on methods to increase silk yield of Bombyxmori (L) ( Lepidoptera : bombycidae ). Ph.D thesis, Tamil Nadu University, Coimbtore, India.
- [19]. RatnaSen, (1988). How does juvenile hormone analogue cause more silk yield. Indian silk: 21-22.
- [20]. Riddiford , L.M (1985). Hormone action at Cellular and Molecular actions of juvenile hormone: general considerations for premetamorpic actions. Adv. Insect physiol., 24: 213-214.
- [21]. Sehnal, F. and Rambold, H. (1985). Brain stimulation of juvenile hormone production in insect larvae. Experimentation 41: 684-685.

- [22]. Slama, K. (1971). Insect juvenile hormone analogues ann. Rev. biochem. 40: 1079-1102.
- [23]. Slama, K.; Wimmer, Z. and Romanuk, M. (1978). Juvenile hormone activity of some glycosidicjuvenogens. Hopperstyler's Physiol. Chem. 359: 1407-1412.
- [24]. Slama, K. (1985). Pharmacology of insect juvenile hormones. In: Biochemistry and pharmacology (Eds G.A Kerkutand L.I Gilbert ), vol. 11: 357-394. Pergamon Press, New York.
- [25]. Vitthalrao B. Khyade; Patil, S.B; Khyade, S.V and Bhavane, G.P (2002). Influence of acetone maceratives of Vitisvinifera on the larval parameters of silkworm Bombyxmori (L). Indian journal of comparative animal physiology vol. 20: 14-18.
- [26]. Vitthalrao B. Khyade; Patil, S.B; Khyade, S.V and Bhavane, G.P (2003). Influence of acetone macerative of Vitisvinifera on the economic parameters of silkworm Bombyx mori (L). Indian journal of comparative animal physiology. Vol. 21: 28-32.
- [27]. Wimmer, Z. and Romanauk, M. (1981). The synthesis of biologically active 2-(4- hydroxyl benzyl)-1cyclohexanol derivation. Coll. Czech. Chem. Commun. 46: 2573-2586.
- [28]. Anjang, Tan ; Hiromasa, Tanaka; Toshiki Tamura and Takahiro Shiotsuki (2005). Precocious metamorphosis in transgenic silkworms overexpressing juvenile hormone esterase. Proc. Natl. Acad. Sci. USA Aug 16, 2005; 102(33): 11751 - 11756. http://genepath.med.harvard.edu/~perrimon/papers/201 3\_Zirin\_DevBio\_Ecdysone.pdf