Effects of lindane(δ- isomer) on adrenal glands in mice (Mus musculus)
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ABSTRACT

Lindane is a broad spectrum organochlorine pesticide contains wide range of application such as in crops protection, treatment of lice and others. Lindane given subcutaneously in mice shows decrease in cortisol level and marked regression in zona fasciculata region of adrenal glands.

Keywords: Lindane, Cortisol, Zona Fasciculate, Adrenal Glands.

I. INTRODUCTION

Lindane (1,2,3,4,5,6-hexachlorocyclohexane) is the only stereoisomer with insecticidal potency and has diversified utilization, which includes protection of crops, prevention of insect borne diseases such as malaria, diseases removal of ectoparasites such as lice and mites, and treatment of human pediculosis. The widespread use of insecticides has caused the scientific community and the public at large to consider seriously the effect of these agents as environmental pollutants and their probable effects on wildlife and human health. Lindane organochlorine pesticide extensively employed for public health and agricultural purpose in developing countries. Lindane is a white, crystalline organic solid. Its formula is C₆H₆Cl₆ and has a molecular weight of 290.8 Its melting point is at 112.5°C, boiling point 323°C, water solubility 7.3 mg/L at 25°C, vapour pressure 4.2 × 10⁻⁵ mm Hg at 20°C, 4.4 × 10⁻³ Pa at 24°C (Source HSBD). Lindane is stable to heat, light, air, carbon dioxide and strong acids. Technical HCH is an isomeric mixture that contains mainly five forms differing only by the chlorine atoms orientation (axial or equatorial positions) around the cyclohexane ring. The five principal isomers are present in the mixture in the following proportions: alpha-hexachlorocyclohexane (53%–70%) in two enantiomeric forms ( (+) alpha-HCH and (-) alpha-HCH), betahexachlorocyclohexane (3%–14%), gamma -hexachlorocyclohexane (11%–18%), delta-hexachlorocyclohexane (6%–10%) and epsilon-hexachlorocyclohexane (3%–5%) [1].

Lindane has been reported to induce oxidative stress [2], membrane perturbation [3], functional impairment in blood brain barrier disturbance in glutathione homeostatis [4] and alteration in cyt P450 monooxygenase enzymes [5].
**Aims and objectives:**
1. To study the effects of lindane in adrenal gland (cortisol level) in serum of mice.
2. To study the histological change in adrenal glands in mice.

**Materials and methodology:**

**Animals:**
Adult albino mice weighing 28-35 gm and approximately 8 weeks of age were procured from Animal House Facility of Department of Zoology, Gauhati University, Assam, India. The animals were housed in properly labelled steel mesh plastic cages with solid bottom containing saw dust and maintained under uniform condition of natural photoperiod (12 hr. light and 12 hr dark), relative humidity, 75%-87% and temperature, 27-30°C. Animals were acclimatized to normal environmental conditions in the laboratory for two weeks before use. Standard diet (pellet diet) and water ad libitum were supplied regularly.

**Chemicals:**
Lindane (8-HCH) was being procured from Zenith India Guwahati, Assam, India. The analytical grade alcohol and distilled water was being supplied by the Department of Zoology, Gauhati University. γ-lindane was used for the experiment because it is still in used in the agricultural field in Assam.

**Preparation of experimental doses:**
Two doses 100 mg/kg bw and 50 mg/kg bw of Lindane were prepared and used in the study. Initially two stock solutions were prepared. For high dose (100mg/kg bw), of test chemical (lindane) was prepared by adding 1 ml of ethanol (Analar Grade) and 9 ml of distilled water to 0.1 ml of the above mentioned doses were injected once daily with the help of 1 ml syringe of 29G (Romson syringe) for 7 and 14 days.

**Experimental grouping of animals:**
Twenty four healthy adult mice were weighed and randomly categorized into four groups (n=6) in ten properly labelled separate cages with steel mesh as lid. The cages were labelled as control group, vehicle control group, 50 mg/kg bw and 100 mg/kg bw respectively. 0.1 ml of the above mentioned doses were injected subcutaneously once daily in the morning around 9 am to 10 am by ‘Romson syringe’ (29 G) for 7 and 14 days. After 7 days, 12 mice i.e., 3 mice from the each of the groups were sacrificed for estimation of effect lindane on various parameters, while the rest 12 mice were dissected after 14 days.

**Table 1: Showing treatment schedule**

<table>
<thead>
<tr>
<th>Experiment Group (n=6)</th>
<th>Treatment (mg/kg bw/animal/day)</th>
<th>Volume Administered (ml)</th>
<th>Duration of treatment (Days)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td></td>
<td></td>
<td>7 and 14</td>
</tr>
<tr>
<td>Vehicle control</td>
<td>(Ethanol:water: 1:9 v/v)</td>
<td>0.1</td>
<td>7and 14</td>
</tr>
<tr>
<td>Low dose</td>
<td>50</td>
<td>0.1</td>
<td>7 and 14</td>
</tr>
<tr>
<td>High dose</td>
<td>100</td>
<td>0.1</td>
<td>7 and 14</td>
</tr>
</tbody>
</table>

Albino mice were taken in 4 different groups (6 animals per group) and two groups of mice were treated with different doses of lindane, 50mg/kg bw (considered as low) and 100mg/kg bw (considered as high) respectively for consecutive 7 and 14 days. A control group was maintained without any treatment. A vehicle control was given ethanol:water (1:9 v/v) 0.1 ml. Treatment schedule: 0.1 ml of test chemical was administered subcutaneously to animals daily in the morning hr daily regularly.
Blood and tissue collection:
Blood samples were drawn by using 2 ml Nypro syringe (26G) using cardiac puncture procedure. Approximately 200 µl of blood was collected and kept separately in micro-centrifuge tubes. The blood samples were then subjected to centrifugation (Eppendorf mini spin centrifuge) at 5000 rpm for 15 min to obtain clear serum. The serum was then collected in newly labelled micro-centrifuge tubes and stored at -20°C for estimating cortisol level.

Following proper laboratory procedure animals were sacrificed one batch on the day 7 and 14 respectively. In the first batch of animals after treatment day 7, all animals were taken in to the laboratory from the animal house. Animals were anesthesised with mild chloroform. Then they were placed on dissecting tray and pinned properly. Adrenal gland was located and taken out to petridishes in normal saline. The weights of the adrenal glands were taken using the Sartorius electronic balance (0.1 mg sensitivity). The adrenal gland transferred to Bouin’s fluid for histology.

Acute toxicity:
The oral LD50 of mice is 86 mg/kg [6]. The acute dermal LD50 of mice is 896 mg/kg.

Cortisol hormone assay:
By electrochemiluscence technique in “Apex diagnostics, Guwahati –781005, Assam”, cortisol hormone assay was done.
**Histopathological study:**
The sample tissues of adrenal gland were kept in Bouin's fluid for 18-24 hours. The fixed specimens were then washed and dehydrated in ascending grades of alcohol (30%, 50%, 70%, 90% and 100%). The specimens were then cleared in xylene, infiltrated and embedded in molten paraffin (60°C), sectioned at 4 micron thickness using microtome (Ernst Leitz Wetzlar GMBH, Germany). The sections were taken in properly affixed slides i.e., fixed with 70% alcohol. The sections were stretched in warm water bath, temperature being maintained at 50-60°C. Proper spreading of the sections was done using a hot plate. The sections were then stained with Haematoxylin and eosin (H & E), and mounted with DPX. The slides were examined under light microscope. Photomicrographs were taken in Microscope (Leica).

**II. Results**

Following are the findings of effect of lindane on adrenal gland

**Table 2:** Effect of different doses of lindane on serum cortisol level. Values are expressed in Mean±SD

<table>
<thead>
<tr>
<th>Experimental group</th>
<th>nmol/L (Mean±SD)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>58.26±3.46</td>
</tr>
<tr>
<td>Vehicle control (7 days)</td>
<td>56.86±3.27</td>
</tr>
<tr>
<td>Vehicle control (14 days)</td>
<td>54.79±2.03</td>
</tr>
<tr>
<td>50 mg/kg bw (7 days)</td>
<td>52.92±2.46</td>
</tr>
<tr>
<td>50 mg/kg bw (14 days)</td>
<td>51.89±1.86</td>
</tr>
<tr>
<td>100 mg/kg bw (7 days)</td>
<td>50.88±1.29</td>
</tr>
<tr>
<td>100 mg/kg bw (14 days)</td>
<td>46.04±1.14</td>
</tr>
</tbody>
</table>

Fig 6: Effect of lindane on the serum cortisol level at 7 days in four animal groups viz., control, 50 mg/kg bw, 100 mg/kg bw and vehicle control. Values expressed in mean ± SD. Values are significant at P <0.05 (* indicates value is significantly different at p <0.05 level compared to the respective control values determined by one way ANOVA analysis).

Fig 7. Effect of lindane on the serum cortisol level at 14 days in four animal groups viz., control, 50 mg/kg bw, 100 mg/kg bw and vehicle control. Values expressed in mean ± SD. Values are significant at P <0.05 (* indicates value is significantly different at p <0.05 level compared to the respective control values determined by one way ANOVA analysis).
Histology:

Fig 8. T.S control adrenal gland 4x magnification

Fig 9. T.S control adrenal gland 10x magnification

Fig 10. T.S low dose adrenal gland 4x magnification

Fig 11. T.S low dose adrenal gland 10x magnification
III. Discussion

The experimental result of the present study indicates that cortisol secretion from the adrenal gland in the treated group decreases which is represented graphically in fig 6. and fig 7. Compared to the low dose of lindane there is marked decrease of cortisol level in high dose. Similar result found by Oskarsson et al. [7], at the highest lindane concentration cortisol secretion was reduced almost to the baseline levels.

It was also observed when the mice were treated with high dose of δ-lindane there is marked regression in the zona fascicula region of the adrenal gland (fig. 12 & 13). In low dose of δ HCH no prominent regression was being observed (fig. 10-11). According to Pulak Lahiri and Sipra Sircar [6] also fasciculata and reticularies zones markedly regressed when treated with γ-lindane.

IV. Conclusion

Pesticides are widely used in agriculture mainly to increase crop yields to cater huge supply of food products for increasing world population as well as to protect crops from pests and control insect-borne diseases. Increased use of pesticides result in contamination of the environment and the excess accumulation of pesticide residues in food products, which has always been a matter of serious concern. Pesticide residues in food and crops are directly related to the irrational application of pesticides to the growing crops. Accumulated pesticide residues in food products have been associated with a broad variety of human health hazards, ranging from short-term effects to long term toxic effects.

The present study firmly established that lindane has many deleterious effects on adrenal glands in mice. Lindane is a persistent organochlorine compound which is widely distributed in the environment. Lindane is considered to be highly toxic. People are exposed to lindane mainly from ingestion of foods contaminated with this pesticide. Additional exposure may come from breathing air contaminated with lindane, dermal contact with contaminated soil or in drinking water. Some people, especially children, may also come into contact with lindane through the use of lotions for scabies or lice control. Infants are also exposed to
lindane and other HCH isomers via their mother’s milk. As from my experiment and secondary sources of data lindane is found to have health hazards effect so its use must be reduced to prevent from its ill effects.

V. Acknowledgement

At the very outset we would like to thank Prof. Jogen Chandra Kalita, Department of Zoology, Gauhati University for his guidance and support during the entire tenure of the experimental period. Sincerely we would also take the opportunity to thank the Department of Zoology, Gauhati University for providing the facilities to perform the experiment.

VI. REFERENCES