

# Green Cellular Delivery of Copper Nanoparticle from Mirabilis Jalapa Flower Extract and Its Antipathogenic Activity

<sup>1\*</sup>C R Rathi, <sup>2</sup> S N Suresh

Department of Biotechnology, Sree Narayana Guru College of Arts and Science, Coimbatore, Tamil Nadu, ,

India

# ABSTRACT

# Article Info

Volume 7, Issue 4 Page Number: 255-263 Publication Issue : July-August-2020 Bio friendly green modest synthesis of nanoparticles are the present research in the extremity of nanotechnology. This study has been undertaken to explore the determinants of copper nanoparticles from 1 mM CuSO<sub>4</sub> solution through profuse concentration of aqueous flower extract of *Mirabilis jalapa* reducing besides immobilizing agent. The attribute of copper nanoparticles was studied by using UV-VIS spectroscopy SEM and XRD. The XRD spectrum of the copper nanoparticles established the presence of elemental copper signal. Green synthesized copper nanoparticles manifests the zone of inhibition against isolated human pathogenic (*Streptococcus species, Bacillus* species, *Staphylococcus species, Klebsiella species* and *E. coli*) bacteria. The analytical chassis contains the flower pigment betalain the natural food dye resources can efficiently use in the production of copper nanoparticles and it could be utilized in various fields in therapeutical and nanotechnology.

Article History

Accepted : 15 Aug 2020 Published : 22 Aug 2020 **Keywords**: Nanoparticles, *Mirabilis jalapa*, Antibacterial activity, UV-VIS spectroscopy, SEM, XRD.

# I. INTRODUCTION

*Mirabilis jalapa* (four O' Clock plant) belongs to the family *Nyctaginacea*e. Mirabilis jalapa bloom all the summer long. They have antifungal, antimicrobial, antiviral, antispasmodic and antibacterial properties (Dimayuga, 1998: Yang, 2001). *Mirabilis jalapa* flower

contains betalain pigments are classified in to red(crimson) betacyanin's and yellow betaxanthins (Strack et al., 2003). Antibacterial properties of copper are documented since 1000 B.C., when copper vessels were used to preserve water (Richard *et al.*, 2002; Castellano *et al.*, 2007). Environmental friendly antimicrobial nano paint can be developed (Kumar *et* 

**Copyright:** © the author(s), publisher and licensee Technoscience Academy. This is an open-access article distributed under the terms of the Creative Commons Attribution Non-Commercial License, which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited

al., 2008). Copper has been known to possess strong antimicrobial properties both in its metallic and nanoparticle forms hence; it has found variety of application in different fields. The FeSO4 attached Ag nanoparticles can be used for the treatment of water and easily removed using magnetic field to avoid contamination of the environment (Gong et al., 2007). The copper nanoparticles are reported to show better wound healing capacity, better cosmetic appearance and scar less healing when tested using an animal model (Tian et al., 2006). Although few copper-containing compounds are approved by the FDA for direct food contact, copper incorporated food packaging is quite widespread in Japan (Appendini and Hotchkiss, 2002). Silica gel micro-spheres mixed with silica thiosulfate are used for long lasting antibacterial activity (Gupta and Silver, 1998) and treatment of burns and various infections (Feng et al., 2000). Copper has been known to possess strong antimicrobial properties both in its metallic and nanoparticle forms hence; it has found variety of application in different fields. The CuSO4attached Cu nanoparticles can be used for the treatment of water and easily removed using magnetic field to avoid contamination of the environment (Gong *et al.*, 2007). Copper nanoparticles can be used for water filtration (Jain and Pradeep, 2005). Use of plant sources offers several advantages such as cost-effectiveness, ecofriendliness and the elimination of high pressure, energy, temperature, and toxic chemicals necessary in the traditional synthesis methods (Sun and Xia, 2002). The most important application of copper and copper nanoparticles is in medical industry such as topical ointments to prevent infection against burns and open wound. Copper ions (Cu+) and its compounds are highly toxic to microorganisms exhibiting strong biocidal effects on many species of bacteria but have a low toxicity towards animal cells (Prema, 2011). It has also been proposed that the antibacterial mechanism of copper nanoparticles is related to the formation of free radicals and

subsequent free radical-induced membrane damage (Danilczuk et al., 2006; Kim et al., 2007). The copper nanoparticle containing poly vinyl nano-fibres also show efficient antibacterial property as wound dressing (Jun et al., 2007). Copper zeolite is used in food preservation, disinfection and decontamination of products (Matsuura et al., 1997; Nikawa et al., 1997). It was also observed that when treated with Cu+, E. coli, a gram-negative bacterium, sustained more structural damages than the gram-positive Staphylococcus aureus (Feng et al., 2000). Silver impregnated medical devices like surgical masks and implantable devices show significant antimicrobial efficacy (Furno et al., 2004). Feng et al., (2000) conducted a study to observe the effects of copper ions on gram positive (Staphylococcus aureus) and gram-negative bacteria (Escherichia coli). Use of plant sources offers several advantages such as effectiveness, eco-friendliness and the elimination of high pressure, energy, temperature, and toxic chemicals necessary in the traditional synthesis methods (Sun and Xia, 2002). Cell membrane detachment from the cell wall, cell wall damage, and electron dense granules outside and, in some instances, inside the cell. It was proposed that condensation of DNA occurred as a protective measure in order to protect the genetic information of the cell (Feng et al.,2000). Copper zeolite is used in food preservation, disinfection and decontamination of products (Matsuura et al., 1997; Nikawa et al., 1997). Copper sulfadiazine depicts better healing of burn wounds due to its slow and steady reaction with serum and other body fluids (Fox and Modak, 1974). Toxicity from copper is observed in the form of argyria, only when there is a large open wound and large amount of copper ions are used for dressing. There are no regular reports of copper allergy (Leaper, 2006). Studies shows that copper nanoparticles anchor to and penetrate the cell wall of Gram-negative bacteria (Morones et al., 2005). cell membrane detachment from the cell wall, cell wall damage, and electron dense granules outside and, in some

instances, inside the cell. It was proposed that condensation of DNA occurred as a protective measure in order to protect the genetic information of the cell (Feng *et al.*, 2000). Copper sulfadazine depicts better healing of burn wounds due to its slow and steady reaction with serum and other body fluids (Fox and Modak, 1974). The main objectives of this study were to (1) Synthesize the copper nanoparticles using aqueous flower extract of Mirabilis jalapa, (2) characterization of copper nanoparticles by using UV-Vis spectroscopy, SEM-XRD (3) analyze antimicrobial properties against human pathogenic bacteria.

# II. MATERIALS AND METHODS

#### i. Sample collection

*Mirabilis jalapa* flower extract were collected from Thodupuzha, Idukki district of Kerala state, India. The fresh flowers were collected and washed with distilled water. The plant materials were thoroughly washed with distilled water and fresh weight were determined. The samples are then oven dried at 50°C for 24 h. The dried samples were powdered using a waring blender and stored in air-tight bottles until further analysis.

#### ii. Extraction Method

*Mirabilis jalapa* flower extract was prepared with 10 g of fresh flower taken in a beaker. It was thoroughly washed with tap water and then with distilled water for at least 2 times and cut into small pieces. The chopped flowers were boiled in 50ml of distilled water for 5 minutes. The flower pigment extract was then cooled and filtered. The filtered flower pigment samples collected and stored in air tight bottles at 4°C until further analysis.

# iii. Synthesis of Copper Nanoparticles

Stock solution was prepared by dissolving 1mM Copper sulphate (CuSO<sub>4</sub>; Merck, Chennai, India) and volume made up to 250 ml with distilled water. 5ml of *Mirabilis jalapa* flower extract was added to 100 ml of 1mM CuSO<sub>4</sub> solution and allowed to react at room temperature.

# iv. Test Microorganisms

The common human pathogenic organisms used (*Staphylococcus, Bacillus, Streptococcus, Salmonella, Klebsiella* and *Escherichia coli*). The test organisms were obtained from microbial stock cultures, District hospital Thodupuzha, Idukki, Kerala.

# v. Characterization of Silver Nanoparticles

UV -Vis Spectroscopy the periodic scans of the optical absorbance between 380 and 500nm with a UV- Vis spectrophotometer (Model 118, Systronics, Mumbai, India) at a resolution of 1 nm were performed to investigate the reduction rate of copper ions by *Mirabilis flower* pigment extract. The reaction mixture was diluted 20 times and used for UV-Vis spectrophotometry. Deionised water was used to adjust the baseline.

#### vi. Antibacterial Assay

Nutrient agar and Muller Hinton agar plates were made according to standard microbiological protocol. Filter paper discs of approximately 6 mm diameter were soaked with 10µl, 20µl, 30µl, 40µl, 50µl respectively of the flower pigment extract, CuSO4 and copper nanoparticle separately and allowed to dry at room temperature for 15 minutes. Muller Hinton Agar plates were prepared and the test microorganisms were inoculated by the spread plate method. Prepared discs were placed in the previously prepared agar plates. Each plate of every test organisms contained discs impregnated with Cu nanoparticle, flower extract, copper sulphate solution and an antibiotic disc. The discs were pressed down to ensure complete contact with the agar surface and distributed evenly so that they were not closer than 24 mm from each other, to Center. The agar plates were then incubated at 37ºC. After 16 to 18 hours of incubation, each plate was examined. The resulting zones of inhibition were uniformly circular with a confluent lawn of growth. The diameters of the zones of inhibition were measured, including the diameter of the disc where the antibiotic was used as control (NCCLS, 1997).



**Figure 1** : a) *Mirabilis jalapa* plant with flower b) *Mirabilis jalapa* fresh flower extract c) Mixture of *Mirabilis jalapa* flower extract and copper sulphate over 2 hours incubation.

## **III. RESULTS AND DISCUSSION**

## *i. Synthesis of Copper Nanoparticles*

After the addition of *Mirabilis jalapa* flower extract to CuSO<sub>4</sub> solution a visible colour change from transparent to dark green was observed which indicates the formation of copper nanoparticle. This occurred due to the reduction of copper ions present in the solution due to terpenoids present in *Mirabilis jalapa* flower extract. After 90 minutes there was no change in the intensity of colour developed, which indicates the completion of reduction reaction. The reduced copper particles are in the range of nano size.

# ii. Characterization of Copper Nanoparticles

*UV Spectrometry* the UV absorption spectrum of copper nanoparticles from *Mirabilis jalapa* flower extract of different concentrations was obtained as given in Figure 2.

# iii. Anti-Bacterial Assay

For *Mirabilis jalapa* flower the zone of inhibition was found to be 13-27 mm for *Klebsiella species*, 17-29 mm for *Bacillus species*, 15-24mm for *E. coli*, 15-26 mm for *Staphylococcus species*, 14-25 mm for *Salmonella species*, and 13-26 mm for *Streptococcus*  species. The study done by Gavane et al., (2012) the zone of inhibition found was 14-26 mm for Klebsiella species. Silver ions and copper salts are used as antimicrobial agents (Russel et al., 1994). However, the high concentrations of copper salts restrict the use of them in present day medicine. Use of metal nanoparticles decreases the concentration of copper and other metal salts. The bactericidal effect of metal nanoparticles has been attributed to their small size and high surface to volume ratio which allows them to interact closely with microbial membranes and is not merely due to release of metal ions in solution or in culture plates (Morones et al., 2005). The mode of action of both copper nanoparticles and copper ions were reported to be similar, although the nanoparticles were reported to be effective at significantly lower concentration than that of the ions (Morones et al., 2005). According to Lok (2006), the attachment of both copper ions and nanoparticles to the cell membrane caused acclimatization of envelope protein precursors causing dissipation of the protein motive force.

#### iv. SEM-XRD Analysis

SEM-XRD analysis proved the effective formation of copper nanoparticles in Mirabilis jalapa flower pigment extract. SEM-XRD analysis was carried out in instrument JSM 6390 with acceleration voltage 20 kV. SEM reveals information about the sample including external morphology, chemical composition and crystalline structure and orientation of materials making up the sample (Yeo SY and Jeong SH (2003). SEM provides detailed high-resolution images of the sample by rastering a focused electron beam across the surface and detecting secondary or back scattered electron signal. The XRD imaging of the copper nanoparticles was performed to confirm the presence of elemental metal signal and provides quantitative information compositional (Oberdorster G. Oberdorster E and Oberdorster J (2005).



**Figure 2:** *Mirabilis jalapa* flower extract UV absorption spectrum of copper nanoparticles.



Figure 3: *Mirabilis jalapa* flower extract copper nanoparticles - SEM microgram of copper nanoparticles.

**Table 1.** Zone of inhibition against various bacteria(Escherichia coli, Salmonella typhi, Staphylococcusspecies, Klebsiella species, Bacillus species,) usingcopper nanoparticles produced by Mirabilis jalapaflower pigment 1/100 dilution.

Microorganis	cont	Sam	Measure zone of				
m	rol	ple	inhibition in mm				
			10	20	30	40	50 µl
			μl	μl	μl	μl	
Escherichia	13	11	15	17	18	21	24
coli							
Salmonella	12	10	14	16	19	23	25
typhi							
Staphylococcu	13	14	15	17	19	20	26
s aureus							
Klebsiella	12	12	13	15	18	22	27
species							
Bacillus	13	11	17	19	22	25	29
species							
Streptococcus	11	14	13	15	16	22	26
species							



**Figure 4** : *Mirabilis jalapa* flower extract copper nanoparticles - X-Ray Diffractogram Cu (Coupled Two Theta/Theta)



Figure 5 : Antimicrobial activity of *Mirabilis jalapa*flower extract green synthesized copper nanoparticles

a) *Streptococcus species*b) *Escherichia coliform*c) *Klebsiella species*d) *Bacillus species*e) *Staphylococcus species*f) *Salmonella species*at the concentration of 10µl, 20µl, 30µl, 40µl, 50µl respectively.

#### **IV. CONCLUSION**

The Mirabilis jalapa flower extract was draw up from fresh flower by boiling it 5 minutes. The prevail extract was of reddish in colour. Freshly prepared flower extract was appended to copper sulphate solution and the reaction takes place at room temperature which resulted in the synthesis of silver The nanoparticles. synthesized nanoparticles UV-VIS nanoparticles were characterized bv spectrometry, SEM and XDX measurements. The UV-VIS spectra of copper nanoparticles formed in the reaction media has absorbance peak at 380 nm. It has been indicated that Mirabilis jalapa extract is competent of fabricating copper nanoparticles that shows good firmness in solution. The synthesized silver nanoparticles were distinguished by UV-VIS spectrum, SEM and XDX measurements. This green synthesis method is substitute to chemical method, since it is contemptible, non-polluting and greenfriendly. The results showed that Mirabilis jalapa flower plays a salient role in the reduction and stabilization of copper to copper nanoparticles. Further, these synthesized silver nanoparticles from *Mirabilis jalapa* shows antibacterial activity on human pathogenic bacteria.

# V. ACKNOWLEDGMENT

The authors are grateful for the cooperation of the Sree Narayana Guru College of Arts and Sciences for necessary support. We also thank those agronomist and laboratory technicians for proving the majority of samples used in the study.

#### VI. REFERENCES

- Appendini P and Hotchkiss JH (2002). Review of antimicrobial food packaging. Innovative Food Science & Emerging Technologies 3(2)113-126.
- [2]. Bhatti, U. and Hanif. M. 2010. Validity of Capital Assets Pricing Model. Evidence from KSE-Pakistan. European Journal of Economics, Finance and Administrative Science, 3 (20). A comparative study of the cytotoxicity of silverbased dressings in monolayer cell, tissue explant, and animal models. Wound Repair and Regeneration 15(1) 94-104.
- [3]. Boateng JS, Matthews KH, Stevens HN and Eccleston GM (2008). Wound healing dressings and drug delivery systems: a review. Journal of Pharmaceutical Sciences 97(8) 2892-291.
- [4]. Castellano JJ, Shafii SM, Ko F, Donate G, Wright TE, Mannari RJ, Payne WG, Smith DJ and Robson MC (2007). Comparative evaluation of silver-containing antimicrobial dressing and drugs. International Wound Journal 4(2) 144-22.
- [5]. Danilczuk M, Lund A, Saldo J, Yamada H, Michalik J (2006). Conduction electron spin resonance of small silver particles. Spectrochimaca Acta Part A: Molecular and Biomolecular Spectroscopy 63(1) 189 - 91.

- [6]. Dimayuga, R E (1998). Antimicrobial activity of medicinal plants from Baja California Sur/Mexico. Pharmaceutical Biol 36:33-43.
- [7]. El-Rafie MH, Mohamed AA, Shaheen TI and Hebeish A (2010). Antimicrobial effect of silver nanoparticles produced by fungal process on cotton fabrics. Carbohydrate Polymers 80(3) 779-782. Feng QL, Wu J, Chen GQ, Cui FZ, Kim TN and Kim JO (2000).
- [8]. Fox CL and Modak SM (1974). Mechanism of silver sulfadiazine action on burn wound infections. Antimicrobial Agents and Chemotherapy 5(6) 582–588.
- [9]. Furno F, Morley KS, Wong B, Sharp BL, Arnold PL and Howdle SM (2004). Silver nanoparticles and polymeric medical devices: a new approach to prevention of infection? Journal of Antimicrobial Chemotherapy 54(6) 1019–1024.
- [10]. Gavhane AJ, Padmanabhan P, Kamble SP and Jangle SN (2012). Synthesis of silver nanoparticles using extract of Neem leaf and Triphala and evaluation of their antimicrobial activities. International Journal of Pharma and Bio Sciences 3(3) 88-100.
- [11]. Gong P, Li H, He X, Wang K, Hu J, Tan W, Zhang S and Yang X (2007). Preparation and antibacterial activity of Fe3O4@Ag nanoparticles. Nanotechnology 18(28) 604-611.
- [12]. Guggenbichler JP, Boswald M, Lugauer S and Krall T (1999). A new technology of micro dispersed silver in polyurethane induces antimicrobial activity in central venous catheters. Infection 27(1) 16-23.
- [13]. Gupta A and Silver S (1998). Silver as a biocide: will resistance become a problem? Nature Biotechnology 16(10) 888. Hussain SM, Hess KL, Gearhart JM, Geiss K and Schlager JJ (2005). In vitro toxicity of nanoparticles in BRL 3A rat liver cells. Toxicology In Vitro 19(7) 975-986.

- [14]. Ip M, Lui SL, Poon VKM, Lung I and Burd A (2006). Antimicrobial activities of silver dressings: an in vitro comparison. Journal of Medical Microbiology 55(1) 59-63.
- [15]. Jain P and Pradeep T (2005). Potential of silver nanoparticle-coated polyurethane foam as an antibacterial water filter. Biotechnology and Bioengineering 90(1) 59-63.
- [16]. Jia J, Duan YY, Wang SH, Zhang SF and Wang ZY (2007). Preparation and characterization of antibacterial silver-containing nanofibers for wound dressing applications. Journal of US-China Medical Science 4(2) 52–54.
- [17]. Kim JS, Kuk E, Yu KN, Kim JH, Park SJ, Lee HJ, Kim SH and Cho MH (2007). Antimicrobial effects of silver nanoparticles. Nanomedicine: Nanotechnology, Biology and Medicine 3(1) 95-101.
- [18]. Klueh U, Wagner V, Kelly S, Johnson A and Bryers JD (2000). Efficacy of silver coated fabric to prevent bacterial colonization and subsequent devicebased biofilm formation. Journal of Biomedical Materials Research Part B: Applied Biomaterials 53(6) 621-631.
- [19]. Kollef MH, Afessa B, Anzueto, A, Veremakis C, Kerr KM, Margolis BD, Craven DE, Roberts PR, Arroliga AC, Hubmayr RD, Restrepo MI, Auger WR and Schinner R (2008). Silver coated endotracheal tubes and incidence of ventilatorassociated pneumonia. The Journal of the American Medical Association 300(7) 805-813.
- [20]. Kumar A, Vemula PK, Ajayan PM and John G (2008). Silver-nanoparticle-embedded antimicrobial paints based on vegetable oil. Nature Materials 7(3) 236-41.
- [21]. Leaper DJ and Durani P (2008). Topical antimicrobial therapy of chronic wounds healing by secondary intention using iodine products. International Wound Journal 5(2) 361-68.

- [22]. Li Y, Leung P, Yao L, Song QW and Newton E (2006). Antimicrobial effect of surgical masks coated with nanoparticles. Journal of Hospital Infection 62(1) 58-63.
- [23]. Lok CN, Ho CM, Chen R, He QY, Yu WY, Sun H, Kwong-Hang P, Chiu TJ and Che CM (2006). Proteomic analysis of the mode of antibacterial action of silver nanoparticles. Journal of Proteome Research 5(4) 916-924.
- [24]. Lok CN, Ho CM, Chen R, He QY, Yu WY, Sun H, Tam PK, Chiu J and Che CM (2007). Silver nanoparticles: partial oxidation and antibacterial activities. Journal of Biological Inorganic Chemistry 12(4) 527-534.
- [25]. Matsuura T, Abe Y, Sato K, Okamoto K, Ueshige M and Akagawa Y (1997). Prolonged antimicrobial effect of tissue conditioners containing silver zeolite. Journal of Dentistry 25(5) 373-377.
- [26]. Morones JR, Elechiguerra JL, Camacho A, Holt K, Kouri JB, Ramirez JT and Yacaman MJ (2005). The bactericidal effect of silver nanoparticles. Nanotechnology 16(10) 2346-2353.
- [27]. Nikawa H, Yamamoto Hamada T, Rahardjo MB and Murata Nakaando S (1997). Antifungal effect of zeolite-incorporated tissue conditioner against Candida albicans growth and/or acid production. Journal of Oral Rehabilitation 24(5) 350-357.
- [28]. Oberdorster G, Oberdorster E and Oberdorster J (2005). Nanotoxicology: an emerging discipline evolving from studies of ultrafine particles. Environmental Health Perspective 113(7) 823-939.
- [29]. Olson ME, Harmon BG and Kollef MH (2002). Silver-coated endotracheal tubes associated with reduced bacterial burden in the lungs of mechanically ventilated dogs. Chest Journal 121(3) 863-870.

- [30]. Ong S, Wu J, Moochhala SM, Tan M and Lu J (2008). Development of a chitosan-based wound dressing with improved hemostatic and antimicrobial properties. Biomaterials 29(32) 4323-4332.
- [31]. Richard JW, Spencer BA, McCoy LF, Carina E, Washington J and Edgar P (2002). Acticoat versus silverlon: the truth. Journal of Burns Surgical Wound Care 1(1)11-20.
- [32]. Russell AD, Path FRC, Si FP and Hugo WB (1994). 7 Antimicrobial activity and action of silver. Progress in Medicinal Chemistry 31(1) 351-370.
- [33]. Shankar SS, Rai A, Ahmad A and Sastry M (2004). Rapid synthesis of Au, Ag and bimetallic Au core- Ag shell nanoparticles using neem (Mirabilis jalapa) leaf broth. Journal of Colloid and Interface Science 275(2)496-502.
- [34]. Silvestry-Rodriguez N, Sicairos-Ruelas EE, Gerba CP and Bright KR (2007). Copper as a disinfectant. Reviews of Environmental Contamination and Toxicology 191(1) 23-45.
- [35]. Strack. D., Vogt,T, Schliemann, W. (2003).Recent advances in betalain research. Phytochemistry, 62:247-269.
- [36]. Sun Y and Xia Y (2002). Shape controlled synthesis of gold and copper nanoparticles. Science 298(5601) 2176-2179.
- [37]. Tian J, Wong KK, Ho CM, Lok CN, Yu WY, Che CM, Chiu JF and Tam PK (2007). Topical delivery of silver nanoparticles promotes wound healing. ChemMedChem 2(1) 171-80.
- [38]. Tripathy A, Raichur AM, Chandrasekaran N, Prathna TC and Mukherjee A (2009). Process variables in biomimetic synthesis of silver nanoparticles by aqueous extract of Mirabilis jalapa flowers. Journal of Nanoparticle Research 12(1) 237-246.
- [39]. Wiley BJ, Im SH, Li ZY, Mclellan J, Siekkenen A and Xia Y (2006). Mane uvering the surface plasmon resonance of copper nanostructures

through shape-controlled synthesis. Journal of Physical Chemistry B 110(32) 15666-15675.

[40]. Yeo SY and Jeong SH (2003). Preparation and characterization of polypropylene/copper nanocomposite fibers. Polymer International 52(7)1053-1057.

# Cite this article as :

C R Rathi, S N Suresh, "Green Cellular Delivery of Copper Nanoparticle from Mirabilis Jalapa Flower Extract and Its Antipathogenic Activity", International Journal of Scientific Research in Science and Technology (IJSRST), Online ISSN: 2395-602X, Print ISSN : 2395-6011, Volume 7 Issue 4, pp. 255-263, July-August 2020. Available at : https://doi.org/10.32628/IJSRST207478 doi Journal URL : http://ijsrst.com/IJSRST207478