International Journal of Scientific Research in Science and Technology



Available online at: www.ijsrst.com





Print ISSN: 2395-6011 | Online ISSN: 2395-602X

doi: https://doi.org/10.32628/IJSRST251395

An Indoor Aeromycological Study of Hospitals in Amravati District

R. D. Vaidya, N. H. Shahare

Department of Botany, Brijlal Biyani Science College, Amravati-444605(M.S.), Maharashtra, India

ARTICLEINFO

Article History:

Accepted: 26 July 2025 Published: 25 August 2025

Publication Issue:

Volume 12, Issue 4 July-August-2025

Page Number:

1091-1097

ABSTRACT

In hospitals, airborne fungal contamination can have serious health effects, especially for patients with weakened immune systems. This study examines the diversity and concentration of airborne fungi in indoor environment of two hospitals in Amravati district of Maharashtra state in India. From two rural hospitals general wards, pathology, and pharmacy were surveyed for aeromycological data. The samples were collected in the month of July 2022 to September 2022. Fungal spores were captured by collecting air samples using the settle plate method. The collected samples were cultured and different species were identified through microscopic examination. The results shows that aeromycoflora was abundant in both hospitals indoor environments. Cladosporium cladosporioides showed the highest percentage contribution in both hospital environments, with 41.93% in Hospital I and 68.55% in Hospital II. In contrast, Aspergillus ochraceus had the lowest contribution in Hospital I (17.74%), while Aspergillus flavus had the lowest in Hospital II (17.74%).

Keywords: Aeromycology, Hospital, Amravati, Cladosporium cladosporioide, Aspergillus ochraceus, Aspergillus flavus

I. INTRODUCTION

"Aerobiology as the interdisciplinary science is closely associated with ecology, botany, phenology, palynology, mycology, microbiology, meteorology, climatology, as well as chemistry and physics of the atmosphere" [22]. Aerobiology studies small particles from biological origin in the air [18].

Aeromycology is the branch of aerobiology that studies the dispersion of spores and other fungal elements in indoor and outdoor air, the changes in their concentrations, and the factors that affect those changes[20]. Aeromycology is the study of the intensity of aerial dispersal of organic matter such as pollen and fungal and/or bacterial spores, both indoors and outdoors[7].

Airborne fungal monitoring began in the 19th century and continued throughout the early part of the 20th century [22]. These early studies aimed to measure the levels of airborne fungi in the outdoor environment [24][14][12]. Researchers have long recognized the connection between fungal spore levels in outdoor air and their presence indoors, as well as the health risks associated with exposure to

these spores in both environments [29]. Noble and Clayton (1963) were among the first to demonstrate the presence of fungi in hospital air[26]. Since then, there has been ongoing discussion in the literature about fungi as potential contaminants or biocontaminants that can affect indoor air quality [23] [11] [8][6].

II. METHODS AND MATERIAL

Selection of Area:

This work was carried out at two different primary health centers of district Amravati, Maharashtra, India. Three wards which are general ward, pathology laboratory and pharmacy of the Hospitals were selected for sample collection. The sample was collected in the month of July 2022 to September 2022. Room temperature and humidity of collection sites was recorded.

Plate Exposure or Settle Plate Method:

Plate exposure or settle plate which involves the opening of plate with specific culture media was used for this study [5][9]. This method allows fungi carrying particles to settle on the respective culture

media. prepared plates are exposed for about 10-15 minutes in the different wards. The plates containing potato dextrose agar (PDA) was used for sample collection. The exposed petri plates were brought into the laboratory and incubated at $28 \pm 1^{\circ}$ C for 7 days. After 3rd, 5th and 7th days of incubation the fungal colonies were counted. At the end of 7th days of incubation the fungus was isolated and pure culture was maintained.

For the purpose of identification and microphotography, slides were prepared with lactophenol cotton blue as the standard stain.

Identification of Collected Samples:

Fungal colonies was initially characterized by cultural, morphological characteristics while the fungal isolates were identified on the basis of colony appearance and microscopic examination, morphology of the spore and hyphae as per Fungal Key of Ellis 1971, Barnett HL, Hunter BB (1972).

Percentage contributions of individual species were calculated as per the standard formula:

III. RESULTS AND DISCUSSION

<u>Table 1</u>: The total count of fungal colonies from indoor environment of hospital-1 from July 2022 to September 2022 at temperature 30°C, 32°C and 31°C respectively.

Sr.	Spore Type	Sampling	July	August	September	Total of colonies found in all
no.	Spore Type	Sites				the months
	Cladosporium cladosporioides	Pathology Lab	6	4	1	11
1		Pharmacy	3	6	3	12
		General -ward	2	-	1	3
		Total	11	10	5	26
	Aspergillus ochraceus	Pathology	1	2	4	7
		Lab				/
2		Pharmacy	-	1	2	3
		General –ward	-	-	1	1
		Total	1	3	7	11
	Aspergillus flavus	Pathology	2	2	2	6
3		Lab	_			U
		Pharmacy	7	1	4	12

Sr.	Cmara Trans	Sampling	July August		September	Total of colonies found in all				
no.	Spore Type	Sites				the months				
		General –ward	1	-	1	2				
		Total	10	3	7	20				
	Unidentified	Pathology	_	-	-	_				
		Lab	_			-				
4		Pharmacy	1	-	-	-				
		General –ward	-	-	-	-				
		Total	1	-	-	-				
	Sterile Hyphae	Pathology	_	_	_					
		Lab	_	_	_	1				
5		Pharmacy	2	-	-	2				
		General -ward	2	-	-	2				
		Total	4	_	-	4				
6	Total		27	16	19	62				

<u>Table 2</u>: The total count of fungal colonies from indoor environment of hospital-II from July 2022 to September 2022 at temperature 29°C, 31°C and 31°C respectively.

Sr.	Cmara Truma	Sampling	Taalaa	Angust	Comtombou	Total of all the	
no.	Spore Type	Sites	July	August	September	months	
	Cladosporium	Pathology	15	13	30	58	
		Lab	13	13	50	30	
1	cladosporioides	Pharmacy	16	14	10	40	
	Ciadosporiolaes	General -ward	3	5	3	11	
		Total	34	32	43	109	
	Aspergillus ochraceus	Pathology	1	14	3	18	
		Lab	1	11	J	10	
2		Pharmacy	-	10	12	22	
		General -ward	-	-	-	-	
		Total	1	24	15	40	
		Pathology	5	2	1	8	
		Lab			1	o .	
3	Aspergillus flavus	Pharmacy	2	-	- 2		
		General -ward	-	-	_	-	
		Total	7	2	1	10	
6	Total		42	58	59	159	

<u>Table 3:</u> Percent count of fungal colonies from July 2022 to September 2022 at temperature 30°C, 32°C and 31°C respectively of hospital- I.

Sr.	Cmore Trme	July	August	September	Total	Percent	count	of	fungal
no.	Spore Type				colonies				
1	Cladosporium	40.74%	62.5%	26.31%	41.93%	6			
1	cladosporioides	10.7170	02.5%	20.5170	T1. 207	U			
2	Aspergillus ochraceus	3.70%	18.75%	36.84%	17.74%	6			
3	Aspergillus flavus	37.03%	18.75%	36.84%	32.25%	6			
4	Unidentified	3.70%	_	-	_				
5	Sterile Hyphae	51.85%	-	-	-				

<u>Table 4:</u> Percent count of fungal colonies from July 2022 to September 2022 at temperature 29°C ,31°C and 31°C respectively of hospital- II.

Sr.	Spore Type	July	August	September	Total	Percent	count	of	fungal
no.	Spore Type				colonies				
1	Cladosporium cladosporioides	80.95%	55.17%	72.88%	68.55%)			
2	Aspergillus ochraceus	2.38%	41.37%	25.42%	25.15%)			
3	Aspergillus flavus	16.66%	3.44%	1.6%	17.74%)			

Indoor aeromycological studies in two different hospitals was conducted in rural areas of Amravati district of Maharashtra, India in July 2022 to September 2022. According to the study's findings, aeromycoflora was abundant in both hospitals indoor environments.

From July to September 2022, a comprehensive assessment of the indoor air quality in Hospital-I was conducted, focusing on the identification of airborne fungal colonies. Over the three-month period, a total of 62 distinct fungal colonies were isolated from exposed Petri plate samples placed at various indoor locations within the hospital. Among these, three were categorized as spore types, four were sterile hyphae indicating fungal growth without visible reproductive structures and one colony remained unidentified due to insufficient morphological features.

Among the identified species, *Cladosporium cladosporioides* was the most dominant, accounting for 41.93% of the total fungal colonies. This species is

a common airborne mold frequently found in indoor environments. *Aspergillus flavus* was the second most prevalent, contributing 32.25% to the total fungal load. This species is significant due to its known allergenic and toxigenic properties, particularly in healthcare settings [30] *Aspergillus ochraceus* had the lowest occurrence, representing only 17.74% of the identified colonies, but its presence is still noteworthy due to its ability to produce ochratoxins, which are harmful to human health[31]. Sterile hyphae shows 51.85% contribution in month July only and unidentified species shows 3.70 % contribution in month July only.

Overall, the findings highlight a seasonal trend in fungal diversity, The predominance of species such as Cladosporium spp. and Aspergillus spp. further emphasizes the need for continuous monitoring and effective indoor air management in hospital environments, particularly to protect immunocompromised patients from potential fungal exposure.

Between July to September 2022, a total of **159 fungal** colonies were isolated from the indoor environment of **Hospital-II**. The monthly distribution of fungal colonies revealed a progressive increase over the three months. This upward trend may reflect seasonal factors such as temperature, humidity, and air circulation, which are known to influence fungal proliferation and spore dispersal in indoor environments.

All identified colonies exhibited three distinct spore types, indicating a range of fungal diversity within the sampled environment.Based on morphological identification from exposed Petri plate samples, Cladosporium cladosporioides emerged as the most dominant fungal species, accounting for 68.55% of the total isolates. This high prevalence aligns with its known adaptability and widespread occurrence in indoor air, particularly in environments with organic material and moisture. In contrast, Aspergillus flavus contributed the lowest proportion, making up only 17.74% of the total colonies. Although less prevalent, A. flavus is of particular concern in hospital settings due to its potential to produce aflatoxins and cause opportunistic infections [21]. Aspergillus ochraceus was moderately represented, comprising 25.15% of the total isolates. This species, while not as dominant as Cladosporium cladosporioides, is notable for its ability to produce ochratoxins, which can pose health risks, especially immunocompromised to individuals[31].

Overall, the data from Hospital-II suggest a relatively high fungal load during the late monsoon months (Rainy season). with *Cladosporium cladosporioides* being the predominant species throughout. The presence of toxigenic *Aspergillus* species, even at lower frequencies, highlights the importance of routine air quality assessments and fungal surveillance in hospital environments to prevent potential health hazards.

Similar to the findings from the "Dr. Manuel Gea Gonzalez" General Hospital, where reported *Cladosporium* spp. as the most frequently isolated

fungal genus in the hospital environment with no significant differences based on sampling time or height [28] .Our study also identified Cladosporium cladosporioides as the dominant airborne fungus in both Hospital-I and Hospital-II. In Hospital-I, it accounted for 41.93% of the total isolates, while in Hospital-II, it was even more prevalent, comprising **68.55%.** These consistent findings across different environments support the idea Cladosporium spp. is a common and persistent component of indoor air, likely due to its environmental adaptability and ability to colonize a wide range of surfaces. The uniformity in its distribution, regardless of sampling variables, further underscores the need for targeted indoor air quality management strategies to limit potential exposure, especially in healthcare settings.

High concentration of airborne spores may lead not only to plant diseases, but often cause skin, eye or nasal irritation and diseases of human respiratory systems, resulting in shortness of breath, alveolitis and asthma [16].

Fungal spores of several genera, such as Cladosporium spp., have been related to asthma exacerbation[20]. Fungus like Aspergillus species can be transmitted from patients or the environment[19]. Aspergillus species may be dangerous for patients of risk group [19]. In those at risk, the microscopic spores can easily enter the upper and lower airways and cause pulmonary aspergillosis[4].

IV. CONCLUSION

According to the current study, in both the hospitals indoor environments contain a variety of aeromycoflora. These research findings could be used to study the impact of airborne fungal spores on human health, which is another significant area of concern. Numerous allergies, airborne illnesses, and respiratory conditions are brought on by exposure to these indoor fungus spores. It might also demonstrate

the direct effects on patients, healthcare professionals, and other people who are in indoor setting.

REFERENCES

- [1]. Asif, A., Zeeshan, M., Hashmi, I., Zahid, U., Bhatti, M,.F Microbial quality assessment of indoor air in a large hospital building during winter and spring seasons. Building and Environment, 68-73135,(2018).
- [2]. Adam, G., Pont, U., Mahdavi, A., Evaluation of thermal environment and indoor air quality in university libraries in vienna. Advance. Material. Research. 899, 315-320,(2014).
- [3]. Awad, A.H., Saeed, Y., Hassan, Y., Fawzy, Y., Osman, M., Air microbial quality in certain public buildings, Egypt: a comparative study. Atmospheric Pollution Research 9(4) DOI:10.1016/j.apr.2017.12.014
- [4]. Anaissie, E. J., Stratton, S. L., Dignani, M. C., Lee, C., Summerbell, R. C., Rex, J. H., Monson, T. P., & Walsh, T. J. Pathogenic molds (including Aspergillus species) in hospital water distribution systems: A 3-year prospective study and clinical implications for patients with hematologic malignancies. Blood, 101(7),2542– 2546(2003). https://doi.org/10.1182/blood-2002-02-0530
- [5]. Bhatia, L., & Vishwakarma, R.. Hospital indoor airborne microflora in private and government-owned hospitals in Sagar City, India. World Journal of Medical Sciences, 5(3), 65–70 (2010).
- [6]. Caillaud, D., Leynaert, B., Keirsbulck, M., Nadif, R., and mould ANSES working group.. Indoor mould exposure, asthma and rhinitis: findings from systematic reviews and recent longitudinal studies. European respiratory review: an official journal of the European Respiratory Society, 27(148). doi:10.1183/16000617.0137-2017(2018)
- [7]. Cariñanos, P., Alcázar, P., Galán, C., Navarro, R., & Domínguez, E. Aerobiology as a tool to help in episodes of occupational allergy in work

- places. Journal of Investigational Allergology & Clinical Immunology, 14(4), 300–308(2004).
- [8]. Dubey, S., Lanjewar, S., Sahu, M., Pandey, K., and Kutti, U. The Monitoring of Filamentous Fungi in the Indoor Air Quality, and Health. Journal of Phytology, 3(4) (2011).
- [9]. Ekhaise, F. O., Ighosewe, O. U., & Ajakpovi, O. D.. Hospital Indoor Airborne Microflora in Private and Government Owned Hospitals in Benin City, Nigeria (2008).
- [10]. Gent, J.F., Ren, P.K., Triche, E., Bracken, M.B., Holford, T.R., Leaderer, B.P., Levelsof household mold associated with respiratory symptoms in the first year of lifein a cohort at risk for asthma. Environ. Health Perspect. 110, 781e786. 2002.
- [11]. Górny, R. L., Reponen, T., Willeke, K., Schmechel, D., Robine, E., Boissier, M., and Grinshpun, S. A.. Fungal Fragments as Indoor Air Biocontaminants. Applied and environmental microbiology, 68(7), 3522–3531 (2002).
- [12]. Hamilton, E. D.. Studies on the air spora. Acta allergologica, 13(2), 143–173 (1959).
- [13]. Hwang, S.H., Cho, J.H.,. Evaluation of airborne fungi and the effects of a platform screen door and station depth in 25 underground subway stations in Seoul, South Korea. Air Qual. Atmos. Health 9, 561e568 (2015).
- [14]. Hirst, J. M.. An automatic volumetric spore trap. The Annals of applied biology, 39(2), 257–265 (1952).
- [15]. Jeon, H.L., Kim, J.H., Cho, T.J., Ji, D.H., Hong, E.J., Son, B.S.,. Assessment of airborne bioaerosols in Korean apartment houses. Quant. Assess. Bioaerosols Apartm. Houses 268e273,(2010).
- [16]. Jędryczka, M.). Aeromycology: Studies of fungi in aeroplankton. Folia Biologica et Oecologica, 10, 18–26. https://doi.org/10.2478/fobio-2014-0013(2014)

- [17]. Long, C.M., Suh, H.H., Catalano, P.J., Koutrakis, P.,. Using time-and size-resolved particulate data to quantify indoor penetration and deposition behavior. Environ.Sci. Technol. 35, 2089e2099, (2001).
- [18]. Kanade, M. B., Wagh, S., Mali, B. S., & Chavan, S. J.. A Review on Aeromycological Studies (2020).
- [19]. Kasdekar, M. M., Duthade, M. M., Damle, A. S., Khapurkuntikar, M. N., Iravane, J. A., Bhakre, J. B., & Gaikwad, A. A.. Air Quality Monitoring of Operation Theaters in Government Medical College and Hospital, Aurangabad, India. International Journal of Current Microbiology and Applied Sciences, 5(6),42–49. https://doi.org/10.20546/ijcmas.2016.506.005(20 16)
- [20]. kasprzyk, i. Aeromycology main research fields of interest during the last 25 years .Annals of agricultural and environmental medicine: AAEM (2008).
- [21]. Klich, M. A.. Aspergillus flavus: The major producer of aflatoxin. Molecular Plant Pathology, 8(6), 713–722 (2007). https://doi.org/10.1111/j.1364-3703.2007.00436.x
- [22]. Maddox, R. L.. On an Apparatus for collecting Atmospheric Particles. The Monthly Microscopical Journal, 3(6), 286–290 (1870).
- [23]. Miller, J. D. Fungi as contaminants in indoor air. Atmospheric Environment, 26(12) (1992)...
- [24]. Morrow, M. B., Lowe, E. P., and Prince, H. E.. Mold fungi in the etiology of respiratory allergic diseases. Journal of allergy, 13(3), 215–226(1941).
- [25]. Myszkowska, D.. Aerobiological studies current state and future challenges. Alergoprofil, 16(1), 8–14(2020). https://doi.org/10.24292/01.AP.161300320
- [26]. Noble, W. C., and Clayton, Y. M. Fungi in the Air of Hospital Wards. Journal of general microbiology, 32, 397–402(1963)..

- [27]. Ponsoni, K., Raddi, M.S.G., Indoor air quality related to occupancy at an airconditioned public building. Braz. Arch. Biol. Technol. 53, 99e103(2010).
- [28]. Ríos-Yuil, J. M., Arenas, R., Fernández, R., Calderón-Ezquerro, M., & Rodriguez-Badillo, R. Aeromycological study at the intensive care unit of the "Dr. Manuel Gea Gonzalez" General Hospital. The Brazilian Journal of Infectious Diseases, 16(5), 432–435 (2012). https://doi.org/10.1016/j.bjid.2012.08.012
- [29]. Richards, M.. Atmospheric mold spores in and out of doors. The Journal of allergy, 25(5), 429–439(1954).
- [30]. Sepahvand, A., Shams-Ghahfarokhi, M., Allameh, A., Jahanshiri, Z., Jamali, M., & Razzaghi Abyaneh, M.. A survey on distribution and toxigenicity of Aspergillus flavus from indoor and outdoor hospital environments. Folia Microbiologica, 56(6), 527–534 (2011). https://doi.org/10.1007/s12223-011-0078-1
- [31]. Skaug, M. A., Eduard, W., & Størmer, F. C.. Ochratoxin A in airborne dust and fungal conidia(2001).
- [32]. Skora, J., Gutarowska, B., Pielech-Przybylska, K., Stepien, L., Pietrzak, K., Piotrowska, M., Pietrowski, P., Assessment of microbiological contamination in the work Environments of museums, archives and libraries. Aerobiologia 31, 389e401(2015).
- [33]. Wu, D., Zhang, Y., Li, A., Kong, Q., Li, Y., Geng, S., Dong, X., Liu, Y., Chen, P.,.Indoor airborne fungal levels in selected comprehensive compartments of the urban utility tunnel in Nanjing, Southeast China. Sustain. Cities Soc. 51(2019).
- [34]. Yazicioglu, M., Asan, A., Ones, U., Vatansever, U., Sen, B., Ture, M., Bostancioglu, M., Pala, O.,. Indoor airborne fungal spores and home characteristics in asthmatic children from Edirne region of Turkey. Allergol. Immunopathol. 109.S56-S56(2004).