

International Journal of Scientific Research in Science and Technology

Available online at : www.ijsrst.com

Print ISSN: 2395-6011 | Online ISSN: 2395-602X



doi : https://doi.org/10.32628/IJSRST

# Deciphering Atorvastatin Calcium Degradation Paradigms : A Review of HPTLC- Facilitated Analytical Methodologies

Ankita Singh<sup>1</sup>, Dr. Ranjeet Kaur Bajwa<sup>2</sup>, Dr. Prafullachandra Tekale<sup>2</sup>, Dr. Gaganjyot Kaur<sup>1</sup>

<sup>1</sup>Department of GNIRD, Guru Nanak Khalsa College of Arts, Science & amp; Commerce (Autonomous), Mumbai 400019, Maharashtra, India

<sup>2</sup>Department of Chemistry, Guru Nanak Khalsa College of Arts, Science & amp; Commerce (Autonomous), Mumbai 400019, Maharashtra, India

## ABSTRACT

Statins are crucial for reducing cholesterol and serve as key examples for understanding degradation dynamics to ensure pharmaceutical efficacy and bioavailability over extended periods. This review highlights the application of high-performance thin-layer chromatography (HPTLC)in elucidating the complex degradation pathways and kinetics of atorvastatin calcium, on stability-indicating methods. Literature shows HPTLC's versatility in resolving degradation products and identifying mechanisms of hydrolysis, oxidation, and photolysis, along with kinetic parameters. The review emphasizes the preparation and validation of HPTLC methods to detect and quantify degradation products during forced degradation studies of atorvastatin calcium, using advanced chromatographic conditions and chemometric tools. It synthesizes research on degradation determinants like temperature, pH, and humidity, and stresses the importance of method validation in terms of specificity, linearity, accuracy, precision, and robustness. Integrating theoretical and practical aspects, this review provides comprehensive insights into the stability of atorvastatin calcium, underscoring the necessity of HPTLC in pharmaceutical quality control.

**Keywords:** Atorvastatin Calcium; High-Performance Thin Layer Chromatography; Degradation dynamic; Stability Indicating Method; Pharmaceutical Quality Control

### INTRODUCTION

Statins are widely used medications that lower cholesterol levels by inhibiting HMG-CoA reductase, a key enzyme in cholesterol biosynthesis in the liver. Elevated cholesterol, especially low-density lipoprotein (LDL) cholesterol, is closely linked to cardiovascular diseases such as coronary artery disease and stroke. By reducing LDL levels, statins significantly lower the risk of atherosclerosis and other cardiovascular complications.[1]

Among statins, atorvastatin calcium is a highly prescribed second-generation drug known for its efficacy in reducing cholesterol and preventing cardiovascular events in high-risk patients, including those with diabetes or hypertension. Given atorvastatin's widespread use, ensuring its stability and effectiveness over its shelf life is vital. Like many pharmaceuticals, atorvastatin is prone to degradation when exposed to environmental stressors such as light, moisture, heat, and oxygen, which can compromise its potency and safety.[2]

High-Performance Thin-Layer Chromatography (HPTLC) plays a critical role in pharmaceutical analysis, particularly for stability-indicating studies. HPTLC offers advantages like high sensitivity, cost-effectiveness, and the ability to resolve multiple compounds, making it an essential tool for identifying and quantifying atorvastatin degradation products. It also facilitates rapid analysis with minimal sample preparation and lower solvent consumption compared to conventional methods, aligning with the growing emphasis on drug stability in the pharmaceutical industry.[3]

In addition to HPTLC, High-Performance Liquid Chromatography (HPLC) is also widely used for atorvastatin degradation studies due to its precision and accuracy. HPLC, known for its high resolution and reproducibility, allows for the separation, identification, and quantification of atorvastatin and its degradation products under various stress conditions. HPLC's versatility in using different detection methods, including UV and mass spectrometry, provides enhanced sensitivity in detecting even trace amounts of degradation products.[4]

The combination of HPTLC and HPLC in atorvastatin stability studies ensures a comprehensive analysis of degradation pathways, contributing to the development of robust formulations and the extension of atorvastatin's shelf life. These techniques are indispensable for ensuring pharmaceutical quality control, regulatory compliance, and patient safety.[5]

## Common Degradation Pathways for Atorvastatin:

Atorvastatin, like many pharmaceutical compounds, is susceptible to degradation through various pathways, which can significantly impact its efficacy and safety. The primary degradation mechanisms include hydrolysis, oxidation, and photolysis:

### 1. Hydrolysis

Hydrolysis is a common degradation pathway where atorvastatin reacts with water, leading to the breakdown of its molecular structure. In acidic or basic conditions, hydrolysis can occur more readily, resulting in the formation of different degradation products. This process is particularly relevant when atorvastatin is exposed to gastrointestinal fluids, which can alter its stability and bioavailability. Hydrolytic degradation can lead to the loss of potency and the generation of inactive or potentially harmful metabolites.[5]

### 2. Oxidation

Oxidation involves the degradation of atorvastatin in the presence of oxygen, often resulting in the formation of various oxidative degradation products. This process can be catalysed by light, heat, or the presence of metal ions. Oxidative degradation can lead to changes in the chemical structure of atorvastatin, impacting its therapeutic effectiveness. Such transformations may generate reactive oxygen species (ROS) that could pose additional risks to patient safety and drug efficacy.[5]

### 3. Photolysis

Photolysis refers to the degradation of atorvastatin when exposed to light, particularly ultraviolet (UV) light. This process can induce the breakdown of the drug's chemical bonds, leading to the formation of degradation products. Photolytic degradation can occur during storage or when the drug is administered,

especially in formulations that are sensitive to light. This pathway is of particular concern in the context of product labelling and storage conditions, as light exposure can compromise the stability and effectiveness of atorvastatin. [5]

## Influence of External Factors on Atorvastatin Degradation

The stability of atorvastatin is significantly influenced by various external factors, including temperature, pH, humidity, and light exposure. Understanding these influences is critical for ensuring the quality and efficacy of atorvastatin in pharmaceutical formulations. Below are the key external factors affecting atorvastatin degradation:

### 1. Temperature

Temperature plays a crucial role in the degradation of atorvastatin. Higher temperatures can accelerate chemical reactions, leading to increased rates of hydrolysis, oxidation, and photolysis. Elevated temperatures can enhance molecular kinetic energy, making degradation pathways more favourable. For instance, storage conditions above recommended levels can result in significant loss of potency due to rapid degradation. Conversely, low temperatures may slow down degradation processes, extending the shelf life of atorvastatin formulations. [6]

## 2. pH

The pH of the environment is another critical factor influencing atorvastatin stability. Atorvastatin is more susceptible to hydrolytic degradation in acidic or basic conditions. For example, under acidic conditions (such as 0.1 M HCl), atorvastatin can undergo significant hydrolysis, resulting in the formation of various degradation products. In contrast, neutral or slightly alkaline conditions may offer more stability, making pH a vital consideration during formulation and storage. The pH-dependent solubility of atorvastatin also affects its bioavailability, necessitating careful formulation design. [6]

## 3. Humidity

Humidity, or the amount of moisture in the environment, can significantly impact atorvastatin stability. High humidity levels can facilitate hydrolysis by providing the necessary water for the degradation process. Moisture can also lead to clumping or degradation of solid dosage forms, potentially affecting the uniformity of the drug within the formulation. Thus, controlling humidity levels during storage and manufacturing is essential for maintaining the integrity of atorvastatin-containing products. [6]

## 4. Light Exposure

Exposure to light, particularly ultraviolet (UV) light, can lead to photodegradation of atorvastatin. Light can induce chemical reactions that break down the drug's molecular structure, resulting in the formation of degradation products that may be less effective or harmful. As a preventive measure, atorvastatin formulations are often packaged in opaque or light-resistant containers to minimize exposure to light during storage and transport. [6]

## 5. Oxygen Levels

Oxygen can contribute to the oxidative degradation of atorvastatin, particularly in the presence of heat or light. Oxidative stress can lead to the formation of free radicals and reactive oxygen species, which can further degrade the drug. To mitigate this risk, formulations may include antioxidants or be packaged in oxygen-impermeable materials to reduce exposure to atmospheric oxygen. [6]

Several key studies have applied HPTLC & HPLC in the degradation analysis of atorvastatin and similar compounds, highlighting its effectiveness in detecting degradation products:

- Dhaneshwar et al. (2007) employed HPTLC to analyse atorvastatin and its degradation products in tablet formulations. The study indicated that atorvastatin was stable under neutral and dry heat conditions but underwent substantial degradation in acidic environments. This work further emphasized the utility of HPTLC in monitoring the degradation behaviour of atorvastatin under various stress conditions. [7]
- Seshachalam and Kothapally (2008) highlighted the application of HPTLC in quantifying atorvastatin in the presence of degradation products. Their findings demonstrated that the method could effectively separate and quantify atorvastatin and its impurities, reinforcing HPTLC's role in quality control and stability testing. [8]
- Shirkhedkar, A. A., & Surana, S. J. (2010) The method validation for atorvastatin calcium via HPTLC involved several tests: Recovery studies used spiked samples at 80%, 100%, and 120%, comparing the results with expected values. Precision was assessed at 400, 500, and 600 ng/band, and robustness tested under varied chromatographic conditions. LOD and LOQ were calculated based on lower concentration ranges. Specificity was confirmed by comparing sample bands to standards. Forced degradation studies under acid, base, hydrogen peroxide, dry heat, and photochemical conditions demonstrated degradation patterns, with samples analysed at 400 ng/band.[9]
- Aiyalu et al. (2011) developed an HPTLC method for the simultaneous analysis of atorvastatin and ezetimibe. Their forced degradation studies revealed significant degradation of atorvastatin under acidic conditions, with up to 89% degradation observed. The method successfully separated atorvastatin from its degradation products, demonstrating HPTLC's capability to assess stability(aiyalu2011). [10]
- Ilango, K., & Kumar, P. S. S. (2013) Forced degradation studies were conducted according to ICH guidelines to assess the stability-indicating properties of the HPTLC method. Methanolic stock solutions of telmisartan (TLM) and atorvastatin (ATV) were refluxed with 0.1 M hydrochloric acid, 0.1 M sodium hydroxide, and 3% hydrogen peroxide at 60°C for 30 minutes. Samples were spotted at concentrations of 120 ng/band for TLM and 30 ng/band for ATV. Significant degradation was observed in acidic (12% TLM, 8% ATV) and alkaline (15% TLM, 28% ATV) conditions, with extra degradation peaks at Rf 0.27 and 0.45 (acid) and 0.42, 0.58, 0.74, 0.80 (alkali). The validated method provided good linearity (r = 0.9998 for TLM, r = 0.9994 for ATV), precision (RSD < 2%), and effective separation of degradation products, confirming its suitability for routine analysis of TLM and ATV in tablets.[11]</li>
- Shah, D. A., Bhatt, K. K., Mehta, R. S., Baldania, S. L., & Gandhi, T. R. (2008) Stress degradation studies on atorvastatin (ATV) and amlodipine (AML) were conducted using acid and alkali hydrolysis, oxidative, and dry heat degradation. Stock solutions (1000 µg/mL) of ATV and AML were treated with 0.1 N NaOH and 0.1 N HCl, heated at 80°C for 1 hour, neutralized, and diluted to a final concentration of 6 µg/mL. Oxidative degradation was performed using 3% hydrogen peroxide under similar conditions. Dry heat degradation involved heating solid drugs at 80°C for 2

354

hours. After degradation, all solutions were analysed using liquid chromatography to investigate interference from degradation products.[12]

Zaheer, Z., Farooqui, M. N., Mangle, A. A., & Nikalje, A. G. (2008) Forced degradation studies on atorvastatin involved acid, base, oxidative, thermal, and photodegradation. For acid and base hydrolysis, atorvastatin tablets were treated with 1 N HCl and NaOH, respectively, heated for 30 minutes, neutralized, diluted with mobile phase, and analysed by HPLC. Oxidative degradation was performed using 3% hydrogen peroxide under similar conditions. Photodegradation was studied by exposing tablets to UV light for 24 hours, and thermal degradation involved heating the drug in a boiling water bath. These treatments revealed degradation products and allowed their separation, ensuring the stability of atorvastatin in various stress conditions.[13]

#### CONCLUSION

The data provided emphasizes the susceptibility of atorvastatin calcium to various degradation pathways, including hydrolysis, oxidation, and photolysis, which significantly impact its stability, efficacy, and safety. The use of sophisticated analytical methods such as High-Performance Thin-Layer Chromatography (HPTLC) and High-Performance Liquid Chromatography (HPLC) is crucial in identifying and quantifying atorvastatin's degradation products under different stress conditions.

HPTLC stands out for its cost-effectiveness, high throughput, and ability to analyze multiple samples with minimal solvent consumption, making it particularly useful for preliminary degradation studies. Meanwhile, HPLC offers superior precision, sensitivity, and the capability to detect even trace amounts of degradation products, making it the gold standard for in-depth analysis. The combination of both techniques provides a robust approach to comprehensively monitor atorvastatin's stability, ensuring pharmaceutical quality control and regulatory compliance.

Environmental factors such as temperature, pH, humidity, light exposure, and oxygen levels play significant roles in atorvastatin's degradation. Managing these factors during formulation and storage is essential for extending the drug's shelf life and preserving its therapeutic efficacy. The cited studies demonstrate that both HPTLC and HPLC are indispensable tools in assessing atorvastatin's degradation under various stress conditions, contributing to better formulation strategies and enhanced drug stability. In conclusion, employing HPTLC and HPLC in concert provides a powerful approach to ensure atorvastatin's stability throughout its shelf life, safeguarding its efficacy and safety for patient use.

#### REFERENCES

- Baldha, R. G., Patel, V. B., & Bapna, M. (2009). Simultaneous spectrophotometric determination of atorvastatin calcium and ezetimibe in tablet dosage form. International Journal of ChemTech Research, 1, 233–236.
- [2] Ballantyne, C. M., Houri, J., & Notarbartolo, A. (2003). Effect of ezetimibe coadministered with atorvastatin in 628 patients with primary hypercholesterolemia. Circulation, 107, 2409–2415. https://doi.org/10.1161/01.CIR.0000068312.51743.BA

- [3] Chaudhari, B. G., Patel, N. M., Shah, P. B., & Patel, L. J. (2007). Stability indicating reversed phase liquid chromatographic method for simultaneous determination of atorvastatin and ezetimibe from their combination drug products. Journal of AOAC International, 90, 1539–1546.
- [4] Prabhu, C., Subramanian, G. S., Karthik, A., Kini, S., Rajan, M. S., & Udupa, N. (2007). Determination of telmisartan by HPTLC—a stability indicating assay. Journal of Planar Chromatography, 20(6), 477– 481.
- [5] International Conference on Harmonization. (2003). Q1A (R2): Stability testing of new drug substances and products. Proceedings of the International Conference on Harmonization, 1–20.
- [6] International Conference on Harmonization. (1996). Q2B: Validation of analytical procedures: Methodology. ICH Secretariat.
- [7] Dhaneshwar, S. S., Dhaneswar, S. R., Deshpande, P., & Patil, M. (2007). Development and validation of a method for simultaneous densitometric estimation of atorvastatin calcium and ezetimibe as the bulk drug and in tablet dosage forms. Acta Chromatographica, 19, 141–148.
- [8] Seshachalam, U., & Kothapally, C. B. (2008). HPLC analysis for simultaneous determination of atorvastatin and ezetimibe in pharmaceutical formulations. Journal of Liquid Chromatography & Related Technologies, 31, 714–721.
- [9] Shirkhedkar, A. A., & Surana, S. J. (2010). Development and validation of a reversed-phase highperformance thin-layer chromatography-densitometric method for determination of atorvastatin calcium in bulk drug and tablets. Journal of AOAC International, 93(3), 798-802.
- [10] Aiyalu, R., & Mani, K. (2012). HPTLC method development, validation, and stress degradation studies for atorvastatin and ezetimibe in multicomponent tablet dosage form. Medicinal Chemistry Research, 21, 1297–1301. https://doi.org/10.1007/s00044-011-9660
- [11] Ilango, K., & Kumar, P. S. S. (2013). Development and validation of stability-indicating HPTLC and HPLC methods for simultaneous determination of telmisartan and atorvastatin in their formulations. Journal of Chemistry, 2013, Article ID 725385, 9 pages. https://doi.org/10.1155/2013/725385
- [12] Shah, D. A., Bhatt, K. K., Mehta, R. S., Baldania, S. L., & Gandhi, T. R. (2008). Stability indicating RP-HPLC estimation of atorvastatin calcium and amlodipine besylate in pharmaceutical formulations. Indian Journal of Pharmaceutical Sciences, 70(6), 754–760. https://doi.org/10.4103/0250-474X.49117
- [13] Zaheer, Z., Farooqui, M. N., Mangle, A. A., & Nikalje, A. G. (2008). Stability-indicating high performance liquid chromatographic determination of atorvastatin calcium in pharmaceutical dosage form. Y.B. Chavan College of Pharmacy, Dr. Rafiq Zakaria Campus, Rauza Bagh, Aurangabad, Maharashtra, India.
- [14] Sonawane, S. S., Shirkhedkar, A. A., Fursule, R. A., & Surana, S. J. (2006). Application of UV spectrophotometry and RP-HPLC for simultaneous determination of atorvastatin calcium and ezetimibe in pharmaceutical dosage forms. Eurasian Journal of Analytical Chemistry, 1, 31–41. https://doi.org/10.1016/j.jpba.2005.07.053
- [15] Erturk, S., Sevinc, E., Erosy, L., & Ficicioglu, S. (2003). An HPLC method for the determination of atorvastatin and its impurities in bulk drug and tablets. Journal of Pharmaceutical and Biomedical Analysis, 33, 1017–1020. https://doi.org/10.1016/S0731-7085(03)00408-4

356

- [16] Godse, V. P., Deodhar, M. N., Bhosale, A. V., Sonawane, R. A., Sakpal, P. S., Borkar, D. D., & Bafana,
  Y. S. (2009). Simultaneous spectrophotometric estimation of ezetimibe and atorvastatin in pharmaceutical dosage form. Asian Journal of Research in Chemistry, 2, 86–89.
- [17] Nilesh, J., Ruchi, J., Hemant, S., Sharad, P., & Deepak Kumar, J. (2010). Spectrophotometric method for simultaneous estimation of simvastatin and ezetimibe in bulk drug and its combined dosage form. International Journal of Pharmacy and Pharmaceutical Sciences, 1, 170–176.
- [18] Rajamanickam, V., Rajasekaran, A., Rathinaraj, B. S., & Anandarajagopal, K. (2010). Development and validation of analytical methods for simultaneous estimation of atorvastatin calcium and ezetimibe in combined dosage form. World Applied Sciences Journal, 9, 1424–1429.
- [19] Lea, A. P., & McTavish, D. (1997). Atorvastatin: A review of its pharmacology and therapeutic potential in the management of hyperlipidaemias. Drugs, 53(5), 828–847.
- [20] Ramsay, L. E., Williams, B., Johnston, G. D., et al. (1999). British hypertension society guidelines for hypertension management. British Medical Journal, 319(7210), 630–635.
- [21] Altuntas, T. G., & Erk, N. (2004). Liquid chromatographic determination of atorvastatin in bulk drug, tablets, and human plasma. Journal of Liquid Chromatography and Related Technologies, 27(1), 83–93.
- [22] Ghosh, C., Jain, I., Gaur, S., Patel, N., Upadhyay, A., & Chakraborty, B. S. (2011). Simultaneous estimation of atorvastatin and its two metabolites from human plasma by ESI-LC-MS/MS. Drug Testing and Analysis, 3(6), 352–362.
- [23] Zarghi, A., Shafaati, A., Foroutan, S. M., & Khoddam, A. (2005). A simple and rapid HPLC method for the determination of atorvastatin in human plasma with UV detection and its application to pharmacokinetic studies. Arzneimittel-Forschung, 55(8), 451–454.
- [24] Palled, M. S., Chatter, M., Rajesh, P. M. N., & Bhat, A. R. (2006). Difference spectrophotometric determination of telmisartan in tablet dosage forms. Indian Journal of Pharmaceutical Sciences, 68(5), 685–686.
- [25] Bahrami, G., Mohammadi, B., Mirzaeei, S., & Kiani, A. (2005). Determination of atorvastatin in human serum by reversed-phase high-performance liquid chromatography with UV detection. Journal of Chromatography B, 826(1-2), 41–45.
- [26] Stanisz, B., & Kania, Ł. (2006). Validation of HPLC method for determination of atorvastatin in tablets and for monitoring stability in solid phase. Acta Poloniae Pharmaceutica, 63(6), 471–476.
- [27] Rao, R. N., Prasad, K. G., Naidu, C. G., & Maurya, P. K. (2011). Development of a validated liquid chromatographic method for determination of related substances of telmisartan in bulk drugs and formulations. Journal of Pharmaceutical and Biomedical Analysis, 56(3), 471–478.
- [28] Farahani, H., Norouzi, P., Beheshti, A., Sobhi, H. R., Dinarvand, R., & Ganjali, M. R. (2009). Quantitation of atorvastatin in human plasma using directly suspended acceptor droplet in liquidliquid-liquid microextraction and high-performance liquid chromatography-ultraviolet detection. Talanta, 80(2), 1001–1006.
- [29] Rao, R. N., Sen, S., Nagaraju, P., Reddy, V. S., Krishnamurthy, P. R., & Bhaskar, S. U. (2006). HPLC determination of telmisartan in bulk and pharmaceutical formulations. Asian Journal of Chemistry, 18(2), 775–782.

- [30] Bing, G. P. Y. L., Xiao, Y. D. W., & Wang, X. (2006). Determination of telmisartan in human plasma using LC-MS and the concentration of pharmacokinetics and bioavailability study. Chinese Journal of Clinical Pharmacy, 13(4), 200–203.
- [31] Hempen, C., Gläsle-Schwarz, L., Kunz, U., & Karst, U. (2006). Determination of telmisartan in human blood plasma—part II: Liquid chromatography-tandem mass spectrometry method development, comparison to immunoassay, and pharmacokinetic study. Analytica Chimica Acta, 560(1-2), 41–49.
- [32] Patil, K. R., Rane, V. P., Sangshetti, J. N., & Shinde, D. B. (2008). A stability-indicating LC method for the simultaneous determination of telmisartan and ramipril in dosage form. Chromatographia, 67(7-8), 575–582.
- [33] Prabhu, C., Subramanian, G. S., Karthik, A., Kini, S., Rajan, M. S., & Udupa, N. (2007). Determination of telmisartan by HPTLC—a stability indicating assay. Journal of Planar Chromatography, 20(6), 477– 481.
- [34] Patil, U. P., Gandhi, S. V., Sengar, M. R., Rajmane, V. S., & Gandhi, S. V. (2010). A validated densitometric method for analysis of telmisartan and atorvastatin calcium in fixed dose combination. Journal of the Chilean Chemical Society, 55(1), 94–96.
- [35] Manoj, S. C., Abhinav, G., & Chakole, R. D. (2012). Simultaneous determination of atorvastatin calcium and telmisartan in pharmaceutical formulations by reverse phase-high performance liquid chromatography. International Journal of Pharmaceutical Chemistry, 2(1), 1–6.
- [36] Delhiraj, N., Ashok, P., Ravikiran, U., & Abhinandhana, P. (2013). A review of various analytical methods on atorvastatin. Indian Journal of Research in Pharmacy and Biotechnology, 1(6), 786.