

Screening of Low Density Polyethylene Degrading Microflora From Garbage Soil

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ABSTRACT

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Plastics have been widely use as a packing material in the form of low density polyethylene. Continuous accumulation of plastic in the environment can cause threat to humidity and environment. In order to stop the accumulation of plastic and to make the surroundings free from plastic, microbes were isolated from medicinal plant soil, garden soil, sewage water soil, Agricultural Soil. These microbes were screened by Clear Zone Technique using polyethylene glycol to confirm the degradation activity. To check the efficiency of biodegradation, weight method was performed under laboratory condition for 2-4 weeks. Experimental data revealed that *Pseudomonas species* have highest plastic degradation capacityas compared to *S.aureus*, *Bacillus subtilis*, *S. pyogenes*, *E.coli*. in natural and artificial conditions.

Keywords : Low density polyethylene, *P.aeruginosa*

I. INTRODUCTION

Plastics are the synthetic polymers of carbon, hydrogen and oxygen which are derived from petrochemical. About 140 million's tons of synthetic polymers are produced worldwide annually with their utility escalating at a rate of 12% per annum (Shimao, 2001). Each year, an estimated 500 billion to 1 trillion plastic bags are consumed worldwide (Roy *et al.*, 2008). Low density polyethylene is one of the major sources of environmental pollution. Polythene waste is recognized as a major threat for marine life. Sometimes, it could cause intestinal blockage in the fishes, birds and marine mammals (Spear *et al.*, 1995and Denuncio *et al.*, 2011). Microbial degradation of plastic caused by oxidation or

hydrolysis using microbial enzymes that lead to chain cleavage of the high molecular weight into low molecular weight oligomers and monomers by aerobic and anaerobic metabolism. Microbes like *Pseudomonas*, *Streptococcus*, *Staphylococcus*, *Micrococcus*, *Rhodococcus*, *Bacillus*, *Brevibacillus*, *Flavobacterium*, *Nocardia* and *Arthrobacter* and fungal species *Aspergillus niger*, *Aspergillus glaucus*, *Cladosporium*, *Fusarium*, *Mucor*, *Penicillium*, *Phanerochaete* and *Trichoderma* involved in biodegradation of polythene (Kumar *et al.*, 2013; Kathiresan K. 2003; Koutny M. *et al.*, 2006). The biodegradation of LDPE due to the disadvantages of other methods such as cost and pollution. Biodegradation is the ability of microorganism to influence abiotic through physical, chemical or

enzymatic action (A. C. Albertsson *et al.*, 1987; E. Chiellini *et al.*, 2003)

II. MATERIALS AND METHOD

Sample collection :

Garbage soil sample were collected from plastic contaminated site from Akola district i.e. Akot file, Malkapur, Ramdas peth, khadaki. The soil sample were collected at a depth of 3-5cm, in sterile container or paper bag and then air dried at room temperature.

Isolation of soil bacteria:

The soil bacteria were isolated by spread technique (Kathiresan, 2003). The 1 gm of Garbage soil sample was taken and mixed in 9ml of distilled water in a test tube and serially diluted. 0.1ml aliquot of various dilutions was spread on nutrient agar medium (Himedia, Mumbai) by using pipette and incubated at 37°C for 24 hr's.

Identification of soil bacteria:

The identification of bacteria was confirmed on the basis of Cultural and Biochemical Characteristics and identified on the basis of Bergey's Manual of Determinative Bacteriology (Holt et al., 1994).

Screening of polyethylene Degrading Microorganisms by Clear Zone Method (Augusta et al., 1993):

Polyethylene Glycol (PEG) was added in mineral salt agar medium at a final concentration of 0.1%(w/v) respectively and the mixture was sonicated for 1 hr. at 120 rpm in shaker. After this the medium was sterilized at 120°C and pressure for 15 lbs/inch² for 20 min. About 15 ml sterilized medium poured before cooling in each plate. The isolated organism were inoculated at 25-30°C for 2-4 weeks. The organism producing zone of clearance around their colonies were selected for further analysis.

Collection of polythene bags:

Low density polyethylene bags (LDPE) used in this study was collected from Akola market, which were 20 um in thick in nature. For the experiment, LDPE were cut into small strips of 2 cm in diameter and they were sterilized with 70% ethanol, air dried and weighed to check initial weight.

Microbial Degradation of Polythene under Laboratory Condition:

Liquid culture method (Orhan et al., 2004):

The pre-weighed strips of polythene bags were aseptically transferred into the conical flask containing 100 ml of nutrients broth and then inoculated with identified polythene degrading microflora and the another conical flask inoculated with consortia of this microflora to observed the plastic degradation capacity of microflora. These flasks were inoculated at 37°C for 10, 20 and 30 days. After a period of time, the strips were washed in 70% ethanol, air dried and weighed to check the final weight. Finally the weight loss was calculated and compared on the below formula

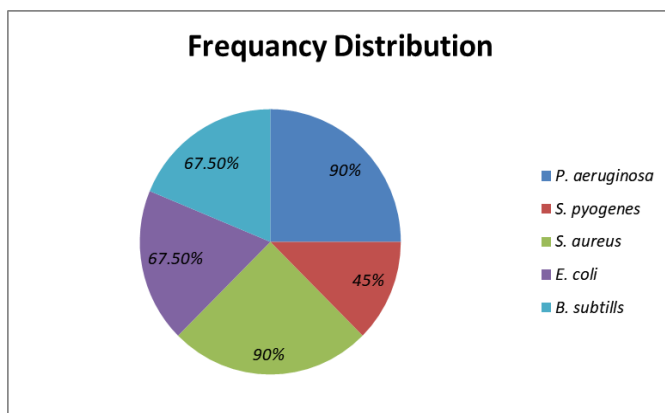
$$\text{weight loss (\%)} = \frac{(\text{Initial weight} - \text{Final weight}) \times 100}{\text{Initial weight}}$$

III. RESULTS AND DISCUSSION

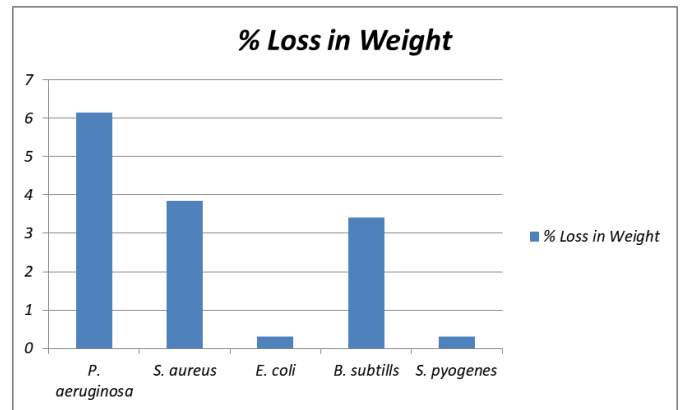
Biodiversity and occurrence of polythene degrading microorganism vary depending on the environment, such as soil, sea compost, activated sludge, etc. It is necessary to investigate the distribution and population of polymer degrading microorganism in various ecosystem. Generally, the adherence of microorganisms on the surface of plastic followed by the colonization of the exposed in the microbial degradation of plastic.

Table 1 : Frequency distribution of Microbial Diversity Obtained from collected samples

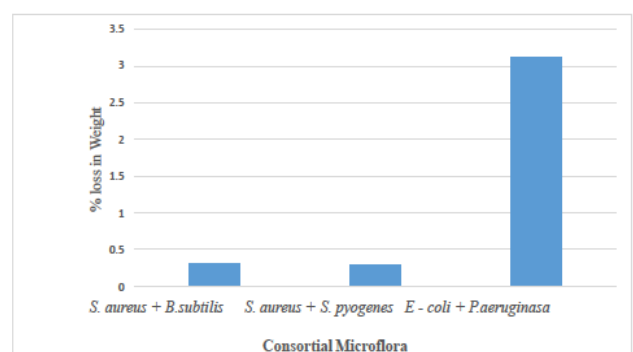
Sr no	Isolates	Frequency distribution	Percentage %
1	<i>P.aeruginosa</i> ,	4	90%
2	<i>Streptococcus.pyogenes</i>	2	45%
3	<i>S.aureus</i>	4	90%
4	<i>E.coli</i>	3	67.5%
5	<i>Bacillus. subtilis</i>	3	67.5%

**Fig 1 :** Frequency distribution of Microbial Diversity Obtained from collected samples - Pie chart

Microorganisms	Initial weight (gm)	Final weight (gm)	Total loss of weight (gm)	% loss in weight
<i>P.aeruginosa</i>	0.0335	0.0323	0.0012	6.15
<i>S.aureus</i>	0.0312	0.0300	0.0012	3.84
<i>E.coli</i>	0.0322	0.0321	0.0001	0.310
<i>B.subtilis</i>	0.0321	0.0310	0.0011	3.42
<i>S.pyogenes</i>	0.0320	0.0319	0.0001	0.312

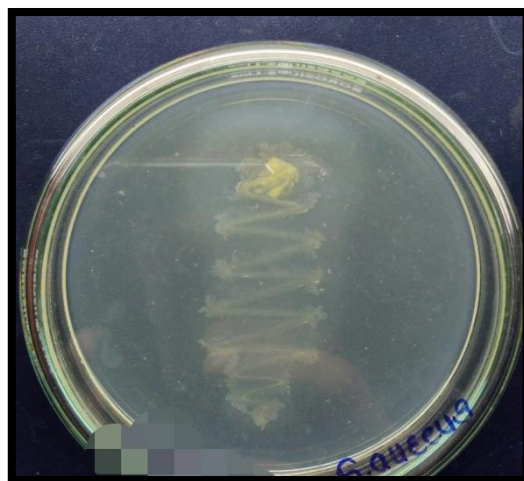
**Fig 2 :** Percentage loss in weight of polythene by the isolated Microflora: Bar diagram.**Table 3 :** % Loss in weight of polythene by the Consortial Microflora

Microorganisms	Initial weight (gm)	Final weight (gm)	Total loss of weight (gm)	% loss in weight
<i>S.aureus + Bacillus subtilis</i>	0.0320	0.0319	0.0001	0.31
<i>S.aureus + S. pyogenes</i>	0.0350	0.0341	0.0009	0.28
<i>E. coli + P.aeruginosa</i>	0.0320	0.0310	0.001	3.125

**Fig 3 :** Percentage loss in weight of polythene by the consortial Microflora: Bar diagram



Polythene bags pieces.



Clear zone formation around the colony on Mineral salt agar



Photograph of Polythene weighing



Biodegradation of Polythene by Flask Method



The present study was aimed to investigate bacterial strain from the plastic contaminated Garbage soil with the ultimate objective of plastic degradation. The sample was collected and studied for its microbial diversity. Microbial diversity observed was

P.aeruginosa, *Streptococcus pyogenes*, *S.aureus*, *E.Coli*, *Bacillus subtilis*.

The soil sample were Collected from the plastic contaminated Garbage soil from the various sites i.e. Akot file, Malkapur, Ramdas peth, khadaki of Akola district. The four soil sample were Collected from the plastic contaminated Garbage soil from the various sites i.e. Akot file, Malkapur, Ramdas peth, khadaki of Akola district. The soil sample were inoculated on various selective media, identified & confirmed on the basis of Bergeys Manual of Determinative Bacteriology. All the isolated bacterial were screen qualitatively for the production of different enzymes such as, Oxidase, Catalase, Gelatinase, Amylase, Urease.

Similar study was reported by Okoh and Atuanya (2014) they reported that *Pseudomonas sp.* Possesses greater potential to degrade polyethylene compared to other bacteria and fungi. Deepika and Jaya Madhuri (2015) also concluded that *Pseudomonas sp.* have significant plastic degradation capacity and it degrade up to 24.22% for the period of 6 months. Similarly, Kyaw *et al.*, (2012) studied that biodegradation of Low Density Polyethylene (LDPE) by *Pseudomonas sp.* They reported that after 120 days of incubation period, the percentage of weight reduction was 20% in *Pseudomonas aeruginosa* (PAO1) strain, 11% in *Pseudomonas aeruginosa* (ATCC) strain, 9% in *Pseudomonas putida* and 11.3% in *Pseudomonas syringae* strain.

The polythene containing mineral salt agar plates were inoculated with the isolated bacteria. All the isolates were screened for their degradation activity. Clear zone was observed after 10 days of incubation at 25-30°C around the colony. On this screening *Pseudomonas sp.*, *Bacillus sp.*, *Streptococcus sp.*, *staphylococcus sp.*, and *E coli* showed high degradation activity. Similar type of findings were

reported by Kathiresan K., (2003) this species of microorganism are associated with the polythene bags and plastic films in soil. Further these soil microorganisms were reported to have the ability for degrading plastics. However the strains with high degradation activity were selected for fermentation.

Determination of weight loss by selected microorganisms were further tested in the laboratory condition to check the ability of degrading polythene and plastics. The bacteria separately allowed to degrade the polythene and plastic incubated for 1 months. After the 1 months period the strips were collected, washed thoroughly using distilled water, shade dried and then weighted to check the final weight. These microorganisms utilize polythene film as a sole source of carbon resulting in partial degradation of plastics. They colonize on the surface of the polyethylene films or plastic cups forming a biofilm. Cell surface hydrophobicity of these organisms was found to be an important factor in the formation of biofilm on the polythene surface, which consequently enhanced biodegradation of the polymers.

Kathiresan and Bingham (2001) reported that bacteria caused the biodegradation ranging from 2.19 to 20.54% for polythene and 0.56 to 8.16% for plastics. Among all the species, *Aspergillus glaucus* was more active than *A.niger* in degrading 28.8% of polythene and 7.26% of plastics within a month. This may be attributed to the thickness of the polythene that is 5-times thinner than the plastics. Once the organisms get attached to the surface, it starts growing by using the polymer as the carbon source. In the primary degradation, the main chain cleaves leading to the formation of low-molecular weight fragments (oligomers), dimers or monomers (Vasile, C., 1993). The degradation due to the extracellular enzymes secreted by the organisms. These low molecular weight compounds are further utilized by the microbes as carbon and energy sources. The

resultant breakdown fragments must be completely used by the microorganisms, otherwise there is the potential for environmental and health consequences (Narayan, R., 2006).

As per the result of study the naturally growing soil microflora like *P.aeruginosa*, *S. aureus*, *E coli*, *S. pyogenes*, *B. subtilis* shows great efficiency in degradation of plastic. *P. aeruginosa* shows maximum percentage loss in the reduction of polythene (6.15 %) and *E. coli* shows minimum percentage loss in the reduction of polythene as (0.310%) compared to isolated microflora i.e the *P. aeruginosa* have a higher degrading effectiveness under laboratory condition. Also we are utilized the consortia of the two different organisms such as *E. coli +P. aeruginosa*, *S.aureus + S. Pyogenes*, *S.aureus+ B. subtilis* to study the degrading efficiency of of isolate, maximum activity was observed in case of consortial organisms i.e. *E coli + P. aeruginosa* (3.125%) results were presented in Table No.1 and Table No. 2 and Graphically represented in Fig. No.1 and Fig. No. 2.

IV. CONCLUSION

The Isolated microbes were native to the site of polyethylene disposal and might show some degradability in natural conditions, yet they also exhibited biodegradation in laboratory conditions on synthetic media. This gives some suggestion that these microbes can be used in both natural and artificial conditions for the purpose of degradation of polymers. Our knowledge, microbes cause greatest degradation of polythene and plastics. Among the microbes *Pseudomonas aeruginosa* having greater degradation ability as compared to *S.aureus*, *Bacillus subtilis*, *S. pyogenes*, *E.coli*. Hence, the further attention is required from microbiologists for commercial degradation and eco-friendly polyethylene with *Pseudomonas aeruginosa*.

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