

# Association of MyCoFlora with Soybean Seed Their Significance and Management

S. V. Aithal<sup>1</sup>, S. S. Patil<sup>2</sup>

<sup>1</sup>Department of Botany, Vai Dhundha Maharaj Deglurkar College, Degloor, Maharashtra, India

<sup>2</sup>Department of Botany, Sharad Chandra ACS College, Naigaon, Maharashtra, India

## ABSTRACT

Seed mycoflora of Soyabean (*Glycine max* L.) seeds were examined in agar plate, blotter method and found the association of fungi i.e., *Aspergillus niger*, *Aspergillus flavus*, *Fusarium oxysporum*, *Fusarium moniliforme*, *Macrophomina phaseolina*, *Sclerotium rolfsii* and *Alternaria alternata*. Seed mycoflora and their culture filtrate caused considered reduction in seed germination and seedling growth. Effect of fungicides, bio-agents and Phyto-extracts on seed mycoflora, germination and vigour index of Soyabean was also evaluated. Seed treatments improved seed germination, vigour index and reduced seed borne mycoflora.

**Keywords:** Soybean, seed mycoflora, seed germination, vigor index.

## I. INTRODUCTION

Soybean (*Glycine max* L) is native of eastern asia. Soybean belongs to the family Fabaceae. It contains 40-44% protein, 20% oil, 8.77% fats and 5.6% fibers. It is also rich in both major and minor minerals.

Major fungal diseases of Soybean (*Glycine max* L) include Leaf spot, Blight, seedling rot, collar rot, charcoal rot, downy mildew (Mukharjee et.al. 1986). Studies on the mycoflora associated with soya bean seeds and their significance have been made by different researchers and they revealed that more than hundred pathogens were known to affect the Soyabean crop (Sinclair, 1982). The fungal pathogens associated with seeds are responsible for several undesirable changes, making them unfit for human consumption as well as sowing (Patil et.al. 2012). The continuous use of chemicals to control diseases results in accumulation of harmful residues of chemicals in the soil, water and seed.

Thus, in recent years, considerable success has been achieved by introducing antagonists to control seed borne fungal pathogens. A notable work has been done for management of seedling diseases of many crops caused by *Rhizoctonia solani* and *Sclerotium rolfsii* both in vitro and in pot culture experiments by using *Trichoderma* (Akhter 1999, Pradeep et al. 2000, Raihan et al. 2003, Haider 2005). Plant extracts also show antifungal activity against wide range of fungi (Abd-Alla et.al 2001). Present investigation was undertaken to find out the fungi associated with the seeds of Soybean, and effect of fungicides, bio-agents and plant-extracts on seed mycoflora, seed germination, seedling length and vigor index.

## II. MATERIALS AND METHODS

### Collection of seed samples: (Source of cultivars)

Seeds of Soybean (*Glycine max* L)-Cv. PK-472, Cv. MAUS-30, Cv. MAUS-38.were obtained from the pulses Researchstation, Marathwada Agricultural University,Badnapur, Jalna, Maharashtra, local farmers,local dealer, etc. The seeds were stored at22°C in cloth bags and used whenever

### Plating of the seed component

Standard blotter paper method and Agar plate method as described by the International Seed Testing Association (ISTA 1996), was used for the isolation of the seed-borne fungi associated with the Soybean seed samples.

### Standard blotter test:

Seeds were equidistantly spaced on moist sterile blotters in Petri plate moist chambers.10 Petri plates of 9” diameter each containing 10 seeds were incubated at 27+2°C for eight days. Observations were made for fungi appearing on seeds every 24 hours and growth was carefully transferred to PDA slants for further studies.

### Agar plating:

Seeds were equidistantly plated on GNA plates aseptically. Colonies which developed during three days were picked up and maintained on PDA/GNA slants. Untreated seeds disinfected externally by treating with10% sodium hypochlorite solution for 10 minutes were used for internally seed mycoflora.

### Seed treatments Treatment with fungicides

Seven chemical fungicides used for seed treatment in different concentration viz; mancozeb 75% WP, carbendazim 50% WP, metalaxyl 8% + mancozeb 64%, pyraclostrobin 5% + metiram 55%, carbendazim 12% + mancozeb 63%, carboxin 75% WP & chlorothalonil 75% WP were evaluated to check their efficacy on germination and vigour index of seeds inoculated with isolated fungi. These treated seeds were evaluated by paper towel method(Khare, M.N.1996)and incubated at  $27 \pm 2$  0 C for 7 days. After end of incubation period observations were recorded as number of germinated seeds, shoot length and root length to calculating vigour index and germination percentage.

### Treatment with Antagonists and Phyto-extracts as bio-priming

Bio-priming was done to study the effect of different bio-agents viz; *Trichoderma viride* @ 0.4%, *Trichoderma harzianum* @ 0.4%, *Pseudomonas fluorescens* @ 5.0 ml, and phyto-extracts viz; Cumin seed extract @ 0.2%, Neem seed extract @ 1.0% & Garlic clove extract @ 1.0% used in different concentration on germination and seedling vigour. Seeds of pigeon pea were soaked in spore suspension of each of the bio-agents and phytoextracts for 24 hours. 2% sugar solution was added into the suspension as sticky material and to provide nutrition to bio-agents which were used for inoculation. Concentration of each bio-agent was adjusted at  $10^7 - 10^8$  cfu/ml with the help hemocytometer and serial dilution technique. Then, effect of respective bio-agents and phyto-extracts were evaluated by Paper towel method (Khare, M.N.1996). Seeds soaked in sterilized distilled water served as control treatment.

### Identification of Fungi:

The various fungi were identified to their generic and specific taxon on the basis of gross colonial and microscopic morphology as per the Key given in "Pictorial atlas of soil and seed fungi: morphologies of cultured fungi and Key to species" (Tsuneo Watanabe, 2002). The fungi were identified on the basis of the shape, measurement and size of the conidiophores, sporangiophores, vesicles, sterigmata, conidia, hyphae, conidial head morphology by using binocular microscope (LABO Bioplan XL). Morphological studies were usually made from the material mounted on slides in lacto phenol and cotton blue. Certain fungi, however particularly those imperfect species which form chains of spores that were easily displaced. Such imperfect fungi were studied without disrupting their growth pattern.

### III. RESULTS AND DISCUSSION

Eight fungi belonging to six genera were isolated by standard blotter paper and agar plate method. Those were *Alternaria alternata*, *Fusarium oxysporum*, *Fusarium moniliforme*, *Sclerotium rolfsii*, *Curvularia lunata*, *Macrophomina phaseolina*, *Aspergillus niger* and *Aspergillus flavus*.

#### Effect of seed infecting fungi on seed health status

Assessment by artificially inoculation of pigeon pea seeds with fungi significantly reduced seed germination, shoot and root length, and thereby seedling vigour index (Table 1). Seeds inoculated with *Fusarium oxysporum* showed lowest seed germination (56%) which was at par with *Fusarium moniliforme* (58%) as earlier reported. All treatments resulted in reduced shoot length, root length and seedling vigor index. *Fusarium oxysporum* recorded minimum shoot length (4.00 cm), root length (5.25 cm) and seedling vigor index (518.20). Treatment with *Aspergillus flavus*, *Fusarium oxysporum*, *Fusarium moniliforme*, *Alternaria alternata*, *Sclerotium rolfsii*, *Curvularia lunata* and *Macrophomina phaseolina* recorded similarly.

#### Effect of culture filtrate of isolated seed infecting fungi on seed health

Seeds treated with culture filtrates of *Aspergillus niger*, *A. flavus*, *Fusarium oxysporum*, *F. moniliforme*, *Alternaria alternata*, *Sclerotium rolfsii*, *Curvularia lunata* and *Macrophomina phaseolina* recorded 43, 45, 54, 58, 62, 66, 69, 73 and 78 % germination, respectively (Table, 2). *Aspergillus niger* recorded minimum shoot length (3.30 cm), root length (4.70 cm) and seedling vigor index (344.20) which was at par with *Aspergillus flavus* 3.45 cm, 4.83 cm and 372.72, respectively. Similarly, *Fusarium oxysporum*, *Fusarium moniliforme*, *Alternaria alternata*, *Sclerotium rolfsii*, *Curvularia lunata* and *Macrophomina phaseolina*, also recorded less shoot length, root length and seedling vigor index. In comparison, significantly highest seed germination (96.00%), shoot length (8.15 cm), root length (12.80 cm) and seedling vigor index (2011.00) were obtained in healthy seeds (Lokesh, M.S. et al 1992).

#### Management Effect of seed treatment with fungicides on soybean seed

Data presented in the (Table 3) revealed significant effect of all fungicides on seed germination, shoot length, root length and seedling vigor index. Seed treated with metalaxyl 8% + mancozeb 64% recorded highest seed germination (93.33%) which was at par with carbendazim 12% + mancozeb 63% (92.00%) and pyraclostrobin 5% + metiram 55% (90.67%). Whereas, mancozeb 75% WP, carbendazim 50% WP, carboxin 75% WP and chlorothalonil 75% WP recorded 84.00, 74.67, 80.00 and 77.33 per cent seed germination, respectively.

Significantly maximum shoot length (11.23 cm) was observed in seed treated with metalaxyl 8% + mancozeb 64% which was at par with carbendazim 12% + mancozeb 63% (11.07 cm). Seeds treated with mancozeb 75% WP (9.20 cm), carbendazim 50% WP (8.30 cm), pyraclostrobin 5% + metiram 55% (9.93 cm), carboxin 75% WP (8.13 cm) and chlorothalonil 75% WP (7.73 cm) also increased shoot length over control. Significantly maximum root length (13.67 cm) was observed in metalaxyl 8% + mancozeb 64% which was at par with pyraclostrobin 5% + mitiram 55% (13.50 cm). Seeds treated with mancozeb 75% WP (11.87 cm), carbendazim 50% WP (10.57 cm), carbendazim 12% + mancozeb 63% (12.87 cm), carboxin 75% WP (9.45 cm) and chlorothalonil 75% WP (9.17 cm) also increased root length over control.

Different bio-agents and phyto-extracts were tested to check their effect on seed germination and seedling health of pigeon pea seeds inoculated with mixture of all isolated fungi. Data presented in the (Table 4) revealed significant effect of all bio-agents and phyto-extracts on seed germination shoot length, root length and seedling vigour index. Seed treated with *Trichoderma viride* recorded highest seed germination (88.00%) which was at par with *Trichoderma harzianum* (85.33%). Whereas, in *Pseudomonas fluorescens*, Neem seed extract, Garlic clove extract and Cumin seed extract recorded 77.33, 73.33, 78.67, 74.67 and 72.00 per cent seed germination, respectively. The results in terms of shoot and root length with seedling vigour index, all the treatments showed larger shoot length, root length and seedling vigour index as compared to control.

#### IV. CONCLUSION

Results of this work show that the three varieties of soybean seeds studied are infected are *Alternaria alternata*, *Fusarium oxysporum*, *Fusarium moniliforme*, *Sclerotium rolfsii*, *Curvularia lunata*, *Macrophomina phaseolina*. and saprophytic fungi are *Aspergillus flavus*, *Aspergillus niger*. The higher seed germination, seedling dry weight, vigour index and lower seed infection are found in fungicides than in bioagent and phyto extracts. From the above discussion it is clear that the studies on seed mycoflora of food crops like Soybean is an important aspect of the plant protection because without seed health tests we cannot touch the target of food security as the healthy seeds are the pre-required of the healthy agriculture.

**Table 1. Effect of seed inoculation with different fungi on seed germination, shoot length, root length and seedling vigour index in pigeon pea.**

Fungi	Seed Germination	Decrease in seed germination over healthy seed	Shoot length (cm)	Decrease in shoot length over healthy seed	Root length (cm)	Decrease in Root length over healthy seed	SVI
<i>Alternaria alternata</i>	76.00	21.65	7.68	18.98	9.78	26.63	1326.30
<i>Fusarium oxysporum</i>	54.00	43.75	4.73	41.96	5.83	54.45	569.70
<i>Fusarium moniliforme</i>	58.00	39.58	5.60	31.28	7.10	44.53	736.90

Sclerotium rolfsii,	69.00	28.12	6.68	18.03	9.65	24.60	1127.10
Curvularia lunata	73.00	73.00	23.96	6.20	23.92	9.15	1121.10
Macrophomina phaseolina	78.00	18.75	7.03	13.74	10.33	19.29	1352.90
Aspergillus niger	43.00	55.20	3.30	59.50	4.70	63.28	344.2
Aspergillus flavus	45.00	53.12	3.45	57.67	4.83	62.27	372.72
Control (Healthy seed)	96.00	-	8.15	-	12.80	-	2011.00

**Table 2. Effect of culture filtrate of isolated seed infecting fungi on seed germination, shoot length, and root length.**

Fungi	Seed Germination	Decrease in seed germination over healthy seed	Shoot length (cm)	Decrease in shoot length over healthy seed	Root length (cm)	Decrease in Root length over healthy seed	SVI
Alternaria alternata	66.00	31.25	5.80	28.83	8.73	31.79	958.50
Fusarium oxysporum	54.00	43.75	4.73	41.96	5.83	54.45	569.70
Fusarium moniliforme	58.00	39.58	5.60	31.28	7.10	44.53	736.90
Sclerotium rolfsii	62.00	35.42	5.30	34.96	6.48	49.37	729.90
Curvularia lunata	69.00	28.12	6.68	18.03	9.65	24.60	1127.10
Macrophomina phaseolina	73.00	23.96	6.20	23.92	9.15	28.51	1121.10
Aspergillus niger	43.00	55.20	3.30	59.50	4.70	63.28	344.20
Aspergillus flavus	43.00	55.20	3.30	59.50	4.70	63.28	344.20
Control (Healthy seed)	96.00	-	8.15	-	12.80	-	2011.00

**Table 3. Effect of seed treatment with fungicides on soybean seed germination, shoot length, root length and seedling vigour index in vitro.**

Treatment	Conc.	Seed Germination	Decrease in seed germination over healthy seed	Shoot length (cm)	Decrease in shoot length over healthy seed	Root length (cm)	Decrease in Root length over healthy seed	SVI
Mancozeb 75% WP,	0.3%	84.00	57.51	9.20	48.39	11.87	61.93	1769.20
Carbendazim 50% WP	0.1%	74.67	40.01	8.30	33.87	10.57	44.20	1409.20
Metalaxyl 8% + Mancozeb 64%,	0.2%	93.33	75.00	11.23	81.12	13.67	86.49	2323.73
Pyraclostrobin 5% + Metiram 55%,	0.2%	90.67	70.02	9.93	60.16	13.50	84.17	2124.53
Carbendazim 12% + Mancozeb 63%,	0.2%	92.00	72.51	11.07	78.55	12.87	75.57	2201.07
Carboxin 75% WP	0.3%	80.00	50.00	8.13	31.13	9.45	28.92	1406.36
Chlorothalonil 75% WP	0.3%	77.33	45.00	7.73	24.68	9.17	25.10	1306.80
Control	-	53.33	-	6.20	-	7.33	-	721.07
CD 0.05%		3.71		0.19		0.20		65.24

## V. REFERENCES

- [1]. Abd-Alla, M.S., Atalla, K.M. and El-Sawi, M.A.M. Effect of some plant waste extracts on growth and aflatoxin production by *Aspergillus flavus*. *Annals Agric. Sci., Ain Shams Univ., Cairo*, 2001; 46: 579-592, 2001.
- [2]. Akhter, N. Biological control of seedling mortality of different crops caused by *Sclerotium rolfsii* using antagonistic fungi. MS Thesis, Department of Plant Pathology, Bangabandhu Sheikh Mujibur Rahman Agricultural University, Gazipur. 76 pp, 1999.
- [3]. Haider, M.M. Biological and chemical control of *Rhizoctonia* dry root rot and foliar blight of soybean (*Glycine max* L. MERR). Department of Plant Pathology, Bangabandhu Sheik Mujibur Rahman Agricultural University, Gazipur. 68 pp, 2005.

- [4] . ISTA. International rules for seed testing. SeedSci. and Techol., 1; 4: 3-49,1996
- [5] . Khare, M.N. Methods to test seeds for associated fungi. Indian Phytopath., 49: 319-328,1996.
- [6] . Lokesh, M.S. and Hiremath, R.V. Studies on seed mycoflora of redgram (*Cajanus cajan* (L.) Millsp.). Karnataka J. Agric. Sci., 5(4): 353-356,1992.
- [7] . Mukherjee K.G. Plant disease of India, Tata Mc Graw Hill Publishing company, New Delhi,1986.
- [8] . Patil, D.P., Pawar, P.V. and Muley, S.M. Mycoflora associated with pigeon pea and chickpea. International Multidisciplinary Research Journal, 2(6): 10-12,2012.
- [9] . Pradeep, K., Anuja, Kumud, K., Kumar, P. and Kumar, K. Bio control of seed borne fungal pathogens of pigeonpea (*Cajanus cajan* (L.) Millsp.). Annals of Plant Protection Sciences 8:30-32,2000.
- [10] . Raihan, M.G., Bhuiyan, M.K.A. and Sultana, N. Efficacy of integration of an antagonist, fungicide and garlic extract to suppress seedling mortality of peanut caused by *Rhizoctonia solani* and *Sclerotium rolfsii*, Bangladesh Journal of Plant Pathology 19 (1&2): 69-73,2003.
- [11] . Sinclair J.B. Compendium of Soybean diseases 2nd edition, American Phytopath Soc. St. Paul Minnesota, USA 41-47pp,1982.